

## RESEARCH ARTICLE

# The Influence of Age and Sex on Genetic Associations with Adult Body Size and Shape: A Large-Scale Genome-Wide Interaction Study



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**Data Availability Statement:** The stratified genome-wide association meta-analysis results for BMI and waist-hip ratio are available from the GIANT Consortium website [www.broadinstitute.org/collaboration/giant](http://www.broadinstitute.org/collaboration/giant)

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## Abstract

Genome-wide association studies (GWAS) have identified more than 100 genetic variants contributing to BMI, a measure of body size, or waist-to-hip ratio (adjusted for BMI,  $WHR_{adjBMI}$ ), a measure of body shape. Body size and shape change as people grow older and these changes differ substantially between men and women. To systematically screen for age- and/or sex-specific effects of genetic variants on BMI and  $WHR_{adjBMI}$ , we performed meta-analyses of 114 studies (up to 320,485 individuals of European descent) with genome-wide chip and/or Metachip data by the Genetic Investigation of Anthropometric Traits (GIANT) Consortium. Each study tested the association of up to ~2.8M SNPs with BMI and  $WHR_{adjBMI}$  in four strata (men  $\leq 50y$ , men  $> 50y$ , women  $\leq 50y$ , women  $> 50y$ ) and summary statistics were combined in stratum-specific meta-analyses. We then screened for variants that showed age-specific effects (G x AGE), sex-specific effects (G x SEX) or age-specific effects that differed between men and women (G x AGE x SEX). For BMI, we identified 15 loci (11 previously established for main effects, four novel) that showed significant ( $FDR < 5\%$ ) age-specific effects, of which 11 had larger effects in younger ( $< 50y$ ) than in older adults ( $\geq 50y$ ). No sex-dependent effects were identified for BMI. For  $WHR_{adjBMI}$ , we identified 44 loci (27 previously established for main effects, 17 novel) with sex-specific effects, of which 28 showed larger effects in women than in men, five showed larger effects in men than in women, and 11 showed opposite effects between sexes. No age-dependent effects were identified for  $WHR_{adjBMI}$ . This is the first genome-wide interaction meta-analysis to report convincing evidence of age-dependent genetic effects on BMI. In addition, we confirm the sex-specificity of genetic effects on  $WHR_{adjBMI}$ . These results may provide



further insights into the biology that underlies weight change with age or the sexually dimorphism of body shape.

## Author Summary

Adult body size and body shape differ substantially between men and women and change over time. More than 100 genetic variants that influence body mass index (measure of body size) or waist-to-hip ratio (measure of body shape) have been identified. While there is evidence that some genetic loci affect body shape differently in men than in women, little is known about whether genetic effects differ in older compared to younger adults, and whether such changes differ between men and women. Therefore, we conducted a systematic genome-wide search, including 114 studies (>320,000 individuals), to specifically identify genetic loci with age- and or sex-dependent effects on body size and shape. We identified 15 loci of which the effect on BMI was different in older compared to younger adults, whereas we found no evidence for loci with different effects in men compared to women. The opposite was seen for body shape as we identified 44 loci of which the effect on waist-to-hip ratio differed between men and women, but no difference between younger and older adults were observed. Our observations may provide new insights into the biology that underlies weight change with age or the sexual dimorphism of body shape.

## Introduction

Body size and shape are independent risk factors for morbidity and mortality [1–6]. They change as people grow older and these changes differ substantially between men and women [7–12]. Subtle sexual dimorphisms are already apparent during early childhood, but differences become more apparent during puberty due, at least in part, to the increasing influence of sex steroid hormones [12–14]. After puberty, sex-differences are largely maintained over the adult life-course. As women age a decline in sex steroid hormones, which coincides with menopause, affects their body shape and composition, resulting in a more android fat distribution [8, 12, 15]. When younger, women tend towards an hourglass body shape with gynoid fat distribution, storing proportionally more fat at thighs and hip than around the waist [12, 16, 17]. At a later age, often after menopause, women's fat storage shifts more upwards around the waist [12, 16, 17]. In men, changes in body fat distribution are subtler than in women, showing a slow but steady increase in waist circumference with age [12]. Thus, after the menopause, the sex-differences in body shape between men and women decrease [12].

This intricate interplay between age and sex on body size and shape is driven by underlying biological processes, involving environmental and genetic factors [7–12, 15]. Elucidating sex- and age-specific genetic effects on body size and shape may provide insights into the biological processes that are involved in the regulation of body weight and fat distribution.

More than 100 genetic loci have been identified for body mass index (BMI), a measure for body size, and for waist-to-hip ratio adjusted for BMI ( $\text{WHR}_{\text{adjBMI}}$ ), a measure of body shape, most of which were identified through our own work in the Genetic Investigation of ANthropometric Traits (GIANT) Consortium [18, 19]. In a recent sex-stratified genome-wide association meta-analysis (up to 133,723 individuals in discovery stage), we searched for variants with sex-specific effects on BMI and  $\text{WHR}_{\text{adjBMI}}$  and identified several loci for which the association

with  $WHR_{adjBMI}$  differed between men and women, whereas no such loci were observed for BMI [10]. However, so far, no GWAS efforts have aimed to identify genetic loci that contribute to differences in body size and shape observed in younger versus older adults, particularly across the menopausal period in women.

We conducted a genome-wide search for loci that exhibit age- and/or sex-specific differences in BMI and  $WHR_{adjBMI}$ . For this, we utilized study-specific genome-wide association statistics separately by sex and by two age groups in each of the studies participating in the GIANT consortium. The two age groups focus on those below and above 50 years of age, as this cut-off coincides with the average age at which women transition through menopause and experience changes in body fat distribution [20–25]. We hypothesize that genetic loci may contribute to the observed differences in body size/shape before age 50y and after age 50y, and that these differences may be sex-specific.

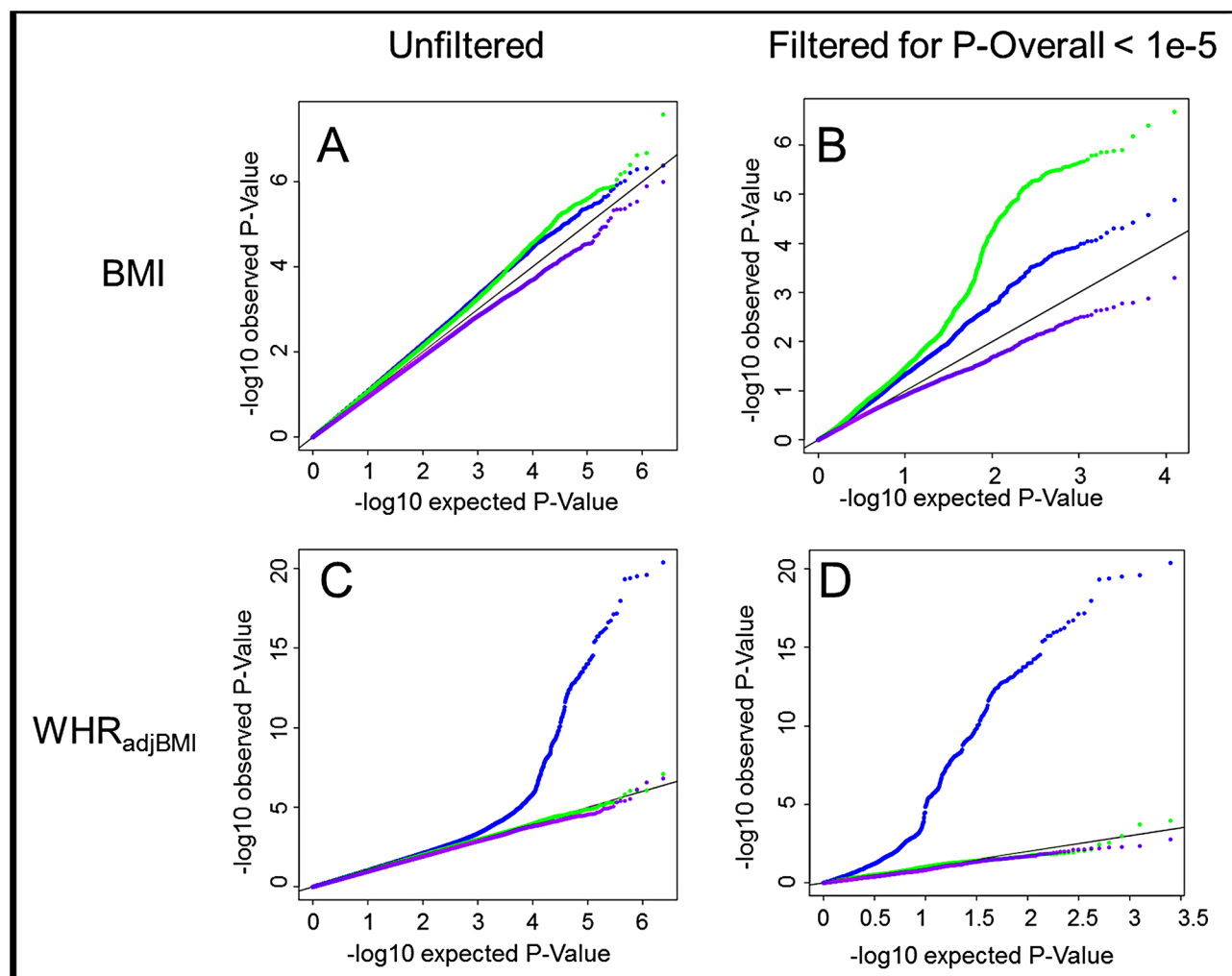
## Results

### Stratified GWAS identifies age- and sex-specific loci for BMI and $WHR_{adjBMI}$

Our total sample comprised up to 320,485 adults ( $\geq 18y$ ) of European ancestry from 114 studies with genome-wide array data imputed to the HapMap reference or genotyped Illumina MetaboChip array data including up to 2.8 million autosomal variants. Details on study-specific analyses, genotyping methods and phenotypic descriptives are given in [S1–S3 Tables](#). To systematically search for genetic loci that influence body size or shape in an age- and sex-specific manner, we first conducted study-specific GWA analyses for BMI and  $WHR_{adjBMI}$  by four strata (men  $\leq 50y$ , men  $> 50y$ , women  $\leq 50y$ , women  $> 50y$ ), and subsequently performed stratified meta-analyses (comprising up to 50,095 men  $\leq 50y$ , 93,201 men  $> 50y$ , 70,692 women  $\leq 50y$ , and 106,497 women  $> 50y$ ) and derived pooled stratum-specific association results ( $P_{men \leq 50y}$ ,  $P_{men > 50y}$ ,  $P_{women \leq 50y}$ ,  $P_{women > 50y}$ ) for each trait. This strategy allowed us to test for three types of interactions: (1) SNPs that demonstrate age-specific effects (SNP  $\times$  AGE,  $P_{agediff}$ ), (2) SNPs that show sex-specific effects (SNP  $\times$  SEX,  $P_{sexdiff}$ ), and (3) SNPs that show age-specific effects that differ between men and women (SNP  $\times$  AGE  $\times$  SEX,  $P_{agesexdiff}$ ). We first performed genome-wide screens using an *a priori* filter; i.e. we examined interaction effects on SNPs that showed evidence of an overall main-effect association ( $P_{Overall} < 10^{-5}$ ). This screen is known to have better power to identify loci with age- or sex-specific effects that are directionally concordant [10, 26]. In a second screen, we examined interaction effects for all SNPs, irrespective of their main-effect association, which allows identification of loci with opposite effect direction in older vs younger adults or in men vs women.

As such, 15 loci with age-specific effects for BMI and 44 loci with sex-specific effects for  $WHR_{adjBMI}$  reached significance after accounting for multiple testing (controlling false-discovery rate, FDR  $< 5\%$ ) ([Figs 1](#) and [S1](#)). No loci were identified with evidence for three way SNP  $\times$  AGE  $\times$  SEX interaction.

In addition to the stratum-specific meta-analyses, we performed (a) a *main effect* meta-analysis that combined the four pooled effect estimates (one from each stratum), providing results for the *overall association* ( $P_{Overall}$ ), assuming effects in age- and sex-groups are the same, and (b) a *joint (main + interaction) meta-analysis approach* ( $P_{joint}$ ) allowing for simultaneous testing of overall association, SNP-by-age and SNP-by-sex interactions [27]. These two screens revealed 83 novel loci of which the association with BMI or  $WHR_{adjBMI}$  reached genome-wide significance ( $P < 5 \times 10^{-8}$ ) ([S2 Fig](#)). This extended discovery is enabled through power augmentation achieved by simultaneously testing main and interaction effects, and/or by accounting for potentially different effects of age and sex on the respective phenotype in the four strata.



**Fig 1. Interaction QQ plots.** Quantile-Quantile plots showing P-Values for age-difference ( $P_{agediff}$ , green), sex-difference ( $P_{sexdiff}$ , blue) and age- and sex-difference ( $P_{agesexdiff}$ , purple). For BMI the P-Values are depicted for all SNPs genome-wide (A) as well as for a limited subset of SNPs that survived pre-filtering on the overall association with BMI,  $P_{Overall} < 1 \times 10^{-5}$  (B). For  $WHR_{adjBMI}$  the P-Values are depicted for all SNPs genome-wide (C) as well as for a limited subset of SNPs that survived pre-filtering on the overall association with  $WHR_{adjBMI}$ ,  $P_{Overall} < 1 \times 10^{-5}$  (D).

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## BMI-novel loci with differential effects in younger and older individuals

Among the 15 loci with significantly different effects (at 5% FDR) on BMI in the younger versus the older individuals, four were novel (near *COBLL1*, *DDC*, *SLC22A3* and *CBLN4*) and 11 were previously established as BMI loci in large-scale *main effect* GWA meta-analyses (near *NEGR1*, *TNNI3K*, *SEC16B*, *TMEM18*, *ADCY3*, *AC016194.1*, *TCF7L2*, *STK33*, *FTO*, *MC4R*, *APOC1*) (S3 Fig and Tables 1 and S4) [19, 28]. Eleven of the 15 age-dependent BMI loci (73%,  $P_{binomial} = 0.06$  for divergence from 50%) showed stronger effects in the younger than in the older group, while the four remaining loci had effects that were more pronounced in the older than in the younger group (Figs 2 and S4). We did not identify BMI-associated loci that showed effects in opposite direction between the younger versus the older group, nor did we find any sex-specific BMI effects. A sensitivity analysis excluding studies with self-report BMI found similar results (S5 Fig).



**Table 1. Fifteen BMI loci showing significant age-differences in adults  $\leq 50$ y compared to adults  $> 50$ y.** The table shows the age-group specific (sex-combined) results, ordered by largest to smallest effect in adults  $\leq 50$ y. All loci were detected by the screen on age-difference that included the a-priori filter on  $P_{Overall} < 10^{-5}$ . The age- and sex-specific results (four strata) and more detailed information on the loci are given in [S4 Table](#).

SNP	Novel Locus <sup>a</sup>	Nearest Gene	Chr	Pos	Alleles <sup>b</sup> EA/OA	EAF	Age $\leq 50$ y			Age $> 50$ y			$P_{Agediff}$
							$\beta$	P	N	$\beta$	P	N	
rs9936385		<i>FTO</i>	16	52376670	C/T	39%	0.093	4.5E-95	115,354	0.073	1.0E-97	197,478	1.6E-04
rs2867125		<i>TMEM18</i>	2	612827	C/T	83%	0.086	6.1E-49	112,934	0.051	2.3E-30	195,579	4.0E-07
rs12955983		<i>MC4R</i>	18	56023969	G/A	28%	0.068	1.7E-41	114,448	0.038	2.0E-23	196,590	6.7E-07
rs6737082		<i>ADCY3</i>	2	24991544	C/A	47%	0.046	6.3E-20	92,191	0.022	5.4E-09	162,112	4.7E-05
rs2821248		<i>NEGR1</i>	1	72348148	A/G	83%	0.042	8.4E-12	106,067	0.017	1.9E-04	188,322	6.2E-04
rs1514174		<i>TNNI3K</i>	1	74765651	C/T	43%	0.039	3.0E-15	92,120	0.012	1.7E-03	161,764	2.8E-06
rs591120		<i>SEC16B</i>	1	176169376	C/G	20%	0.033	4.9E-14	115,337	0.014	2.8E-05	197,481	3.1E-04
rs11908421	yes	<i>CBLN4</i>	20	53813074	T/C	81%	0.033	8.7E-08	92,575	0.007	1.2E-01	162,284	4.3E-04
rs4947644	yes	<i>DDC</i>	7	50586370	T/C	51%	0.030	7.7E-10	91,980	0.009	1.7E-02	158,555	2.5E-04
rs10840060		<i>STK33</i>	11	84566621	C/A	50%	0.029	3.8E-11	110,697	0.011	2.0E-03	187,808	4.0E-04
rs1459180		Intergenic	8	77144822	G/T	58%	0.027	3.1E-09	112,913	0.009	1.6E-02	190,729	6.0E-04
rs17747324		<i>TCF7L2</i>	10	114742493	T/C	77%	0.004	4.8E-01	111,572	0.031	2.6E-13	193,773	4.7E-05
rs3127574	yes	<i>SLC22A3</i>	6	160711360	C/G	51%	0.001	7.9E-01	113,057	0.019	2.3E-08	195,472	6.8E-04
rs3769885	yes	<i>COBLL1</i>	2	165300636	A/G	48%	-0.001	9.1E-01	107,703	0.020	3.9E-09	192,513	1.1E-04
rs4420638		<i>APOC1</i>	19	50114786	A/G	82%	-0.007	3.6E-01	83,196	0.040	8.9E-12	152,014	2.1E-07

Chr: Chromosome; Pos: position; EAF: Effect Allele Frequency; EA: Effect allele; OA: Other allele

<sup>a</sup> 'Yes' if the locus is mentioned as BMI locus for the first time

<sup>b</sup> Effect allele is according to the BMI increasing allele according to the associated sex.

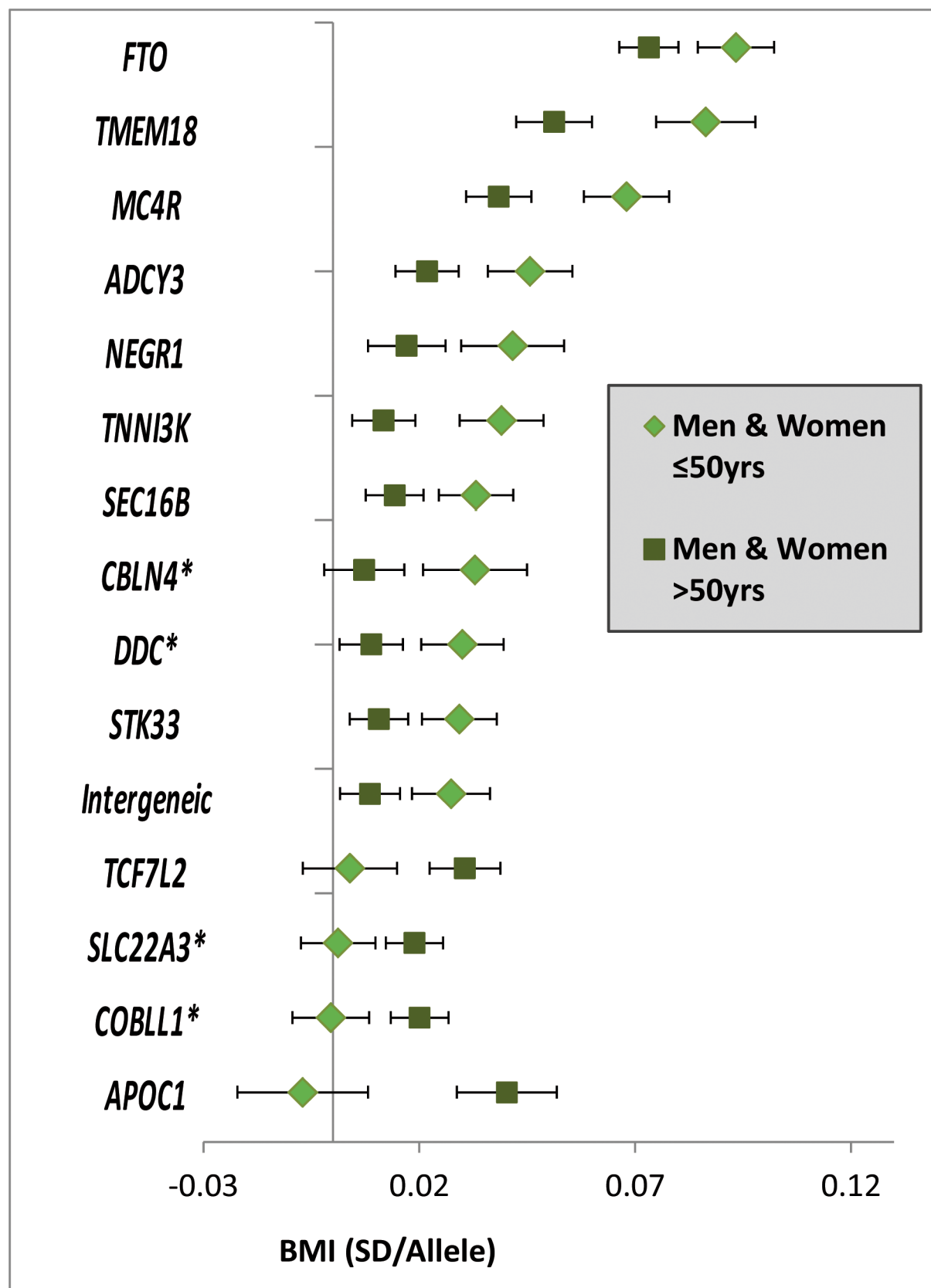
doi:10.1371/journal.pgen.1005378.t001

## WHR<sub>adjBMI</sub>—additional genetic loci contribute to differences between men and women

Unlike for BMI, no WHR<sub>adjBMI</sub>-associated loci with significant difference between the age-groups were observed. Yet, 44 loci showed significantly different effects on WHR<sub>adjBMI</sub> between women and men of which 17 loci were novel (near *TTN*, *IRS1*, *CDH10*, *IQGAP2*, *SIM1*, *ISPD*, *KLF14*, *SGCZ*, *PTPRD*, *RXRA*, *GANAB*, *SLC2A3*, *LEMD3*, *GNPNAT1*, *RPS6KA5*, *CECR2*, *HMGXB4*) and 27 loci had been previously established in *main-effect* GWA meta-analyses for WHR<sub>adjBMI</sub> ([S6 Fig](#) and [Tables 2](#) and [S5](#)). Of the 27 previously established WHR<sub>adjBMI</sub> loci, sex-differences had already been reported for 17 loci [[10](#), [29](#)] [[18](#)]. Our genome-wide screen established sex-specific effects for an additional 10 of the previously established loci with a main-effect on WHR<sub>adjBMI</sub> (near *GORAB*, *LY86*, *ITPR2*, *PIGU*, *EYA2*, *KCNJ2*, *MEIS*, *EYA1*, *CCDC92*, *NSD1*). Of the 44 sex-specific loci, 11 loci showed opposite effect directions in women versus men and 33 showed a significant effect in one and a smaller or no effect in the other sex. Consistent with previous observations, almost all of these 33 loci (28 out of the 33,  $P_{binomial} = 3.3 \times 10^{-5}$ ) showed more pronounced effects in women than in men ([Figs 3](#) and [S7](#)). Again, a sensitivity analysis excluding studies with self-report waist and hip circumference found similar results ([S8 Fig](#)).

## No evidence for loci with simultaneous age- and sex-specific effects

We searched for loci with sex-specific effects on WHR<sub>adjBMI</sub> that differ between the two age-groups and for loci with age-specific effect on BMI that differ between men and women by testing a three-way interaction (SNP x AGE x SEX,  $P_{agesexdiff}$ ). We first tested for this three-way interaction in the 59 SNPs identified with an age-difference (15 loci for BMI) or a sex-difference (44 loci for WHR<sub>adjBMI</sub>), as described above. However, none of these 59 loci showed a



**Fig 2. Age-dependent BMI loci.** Effect estimates (beta  $\pm$ 95CI) per standard deviation in BMI and risk allele for loci showing age-differences in men & women  $\leq$ 50y compared to men & women >50y. Loci are ordered by greater magnitude of effect in men & women  $\leq$ 50y compared to men & women >50y. (95%CI: 95% confidence interval; BMI: body mass index; SD: standard deviation, \*Newly identified loci).

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significant three-way interaction ( $P_{\text{agesexdiff}} > 0.00084 = 0.05/59$ , Bonferroni corrected) ([S4 and S5 Tables](#)). When screening for the three-way interaction genome-wide, no such loci were identified (at 5% FDR) ([Fig 1](#)).

## Detecting loci with age- or/and sex-interaction requires extremely large sample sizes

We analytically computed the statistical power of our screens to identify SNP x AGE, SNP x SEX or SNP x AGE x SEX interaction effects, assuming a total sample size of 300,000 individuals distributed across four equally sized strata and considering a range of effect size configurations informed by previous observations ([S9](#), [S10](#) and [S11 Figs](#)). For example, for a medium genetic effect on BMI ( $R^2 = 0.037\%$  as observed previously for a locus near *MAP2K5* [[28](#)]), our screens had (i) sufficient power to identify genetic loci with two-way SNP x AGE or SNP x SEX interactions (i.e. loci with effect in one stratum and not in the other, so-called *pure two-way interaction*, power = 86%, or loci with effect in both strata, but with opposite effect direction, power = 99%), (ii) sufficient power to detect *extreme three-way interaction* SNP x AGE x SEX, typically involving a biologically-unlikely scenario with opposite effect directions across both AGE and SEX (power = 99%), but (iii) insufficient power to identify loci with biologically more plausible three-way interactions (in the range of  $R^2$  of 0.01–0.05%), i.e., loci that have an effect in only one stratum and not in the other three strata, *1-stratum interaction*, power = 2%, or those with a similar effect in three strata and not in the fourth, *3-strata interaction*, power = 21% ([Fig 4](#)). Identification of loci with medium 1-stratum ( $R^2 = 0.037\%$  in one stratum and  $R^2 = 0$  in the other three strata) or 3-strata ( $R^2 = 0.037\%$  in three strata and  $R^2 = 0$  in one stratum) interaction effects with a power of 80%, would require a total sample size of 750,000 or 600,000 individuals, respectively.

Reducing the multiple testing burden by applying a filter on the overall meta-analysis to first identify SNPs with main effects ( $P_{\text{Overall}} < 10^{-5}$ ) improved the statistical power to identify loci with specific interaction scenarios: (i) loci with pure two-way interaction effects (e.g. 30% power increase to detect SNP x AGE with  $R^2 = 0.037\%$  and  $R^2 = 0$  in the two strata), or (ii) loci with 3-strata interaction effects (e.g. 21% power increase for loci with  $R^2 = 0.037\%$  in three strata and  $R^2 = 0$  in one stratum) ([Figs 4 and S9](#)).

With our sample size of 300,000 subjects and equally sized strata we had 80% power to detect (i) 1-stratum interaction with  $R^2 = 0.09\%$  in one stratum ( $R^2 = 0$  in the other three strata), (ii) 3-strata interaction with  $R^2 = 0.07\%$  in three strata ( $R^2 = 0$  in one stratum), or (iii) pure two-way interaction with  $R^2 = 0.03\%$  in one stratum ( $R^2 = 0\%$  in the other stratum).

In summary, this analysis suggests that our study is sufficiently powered to detect even subtle two-way interaction effects, and would certainly include effect-sizes that would be considered biologically or clinically important. While even more subtle interactions may be occurring, it appears likely that in this effort, we have detected the most important age- and sex- interactions for body size and shape.

## Association of identified loci with other traits

To examine whether the age- and sex-specific effects of the identified BMI and  $\text{WHR}_{\text{adjBMI}}$  loci translate into similar age- and sex-effects on obesity-related cardiometabolic traits, we gathered results from the ICBP, CHARGE and Global-BPGen consortia (age-specific and sex-specific



**Table 2. Forty-four  $WHR_{adjBMI}$  loci showing significant sex-differences.** The table shows the sex-specific (age-group combined) results, ordered by largest, positive effect in women to largest, negative effect in women. The age- and sex-specific results (four strata), more detailed information on the loci and on the screens for which they were detected are given in [S5 Table](#).

SNP	NovelLocus <sup>a</sup>	Novel Sexdiff <sup>b</sup>	NearestGene	Chr	Pos	Alleles <sup>c</sup> EA/OA	Women				Men			
							EAF	$\beta$	P	N	$\beta$	P	N	$P_{Sexdiff}$
rs2820443			LYPLAL1	1	217820132	T/C	72%	0.063	5.2E-36	111,691	0.000	9.8E-01	93,780	1.1E-18
rs998584			VEGFA	6	43865874	A/C	48%	0.060	3.1E-32	109,533	0.015	5.0E-03	87,177	5.0E-10
rs6717858			COBLL1	2	165247907	T/C	59%	0.054	2.7E-31	110,110	-0.009	7.2E-02	90,259	4.2E-21
rs4616635			ADAMTS9	3	64677315	C/G	72%	0.049	4.0E-23	114,021	0.008	1.5E-01	93,679	7.5E-09
rs2811434			PLXND1	3	130822305	T/G	79%	0.046	8.8E-14	91,914	-0.005	4.7E-01	66,742	3.0E-08
rs1936811	yes		RSPO3	6	127425553	T/A	61%	0.043	5.6E-17	91,862	0.015	1.4E-02	67,436	2.0E-04
rs10743579			ITPR2	12	26352412	A/C	25%	0.043	1.4E-13	91,035	0.020	2.3E-03	67,178	8.7E-03
rs6958350			NFE2L3	7	25838458	T/C	25%	0.038	4.8E-14	114,759	0.016	4.9E-03	91,294	2.1E-03
rs1443512			HOXC13	12	52628951	A/C	24%	0.038	3.0E-13	114,486	0.016	5.8E-03	88,811	3.8E-03
rs7830933			NKX2-6	8	23659269	A/G	77%	0.037	4.4E-13	116,052	-0.004	5.3E-01	93,504	4.8E-08
rs11057396	yes		CDC92	12	122985015	A/C	67%	0.037	1.2E-10	78,489	0.005	4.6E-01	53,789	2.6E-04
rs1294404	yes		LY86	6	6680021	A/G	61%	0.035	3.0E-14	116,324	0.016	1.4E-03	92,668	4.3E-03
rs9687846			MAP3K1	5	55897651	A/G	19%	0.035	1.9E-09	116,005	0.000	9.8E-01	93,710	3.5E-05
rs6018158	yes		EYA2	20	44971841	T/C	41%	0.033	7.5E-11	93,476	0.012	3.9E-02	67,612	5.0E-03
rs745578	yes		EYA1	8	72628878	A/G	24%	0.033	2.9E-08	92,963	0.010	1.6E-01	67,179	9.3E-03
rs1045241			TNFAIP8	5	118757185	C/T	71%	0.033	4.8E-11	116,314	0.000	9.3E-01	93,754	3.4E-06
rs7492628	yes		RPS6KA5	14	90616889	G/C	30%	0.031	2.3E-08	91,645	0.007	2.5E-01	66,029	3.9E-03
rs17819328			PPARG	3	12464342	G/T	43%	0.031	8.5E-11	109,626	0.004	4.3E-01	88,650	7.9E-05
rs12443634			CIMP	16	80081775	A/C	29%	0.031	3.2E-08	93,188	-0.009	1.8E-01	66,051	2.4E-06
rs4656767	yes		GORAB	1	168646351	A/C	71%	0.029	5.3E-09	115,682	0.006	2.8E-01	91,023	1.3E-03
rs13029520	yes		MEIS1	2	66626466	T/C	40%	0.028	3.5E-08	86,851	0.007	2.1E-01	62,091	6.9E-03
rs2092029	yes		HMGXB4	22	33982241	C/T	33%	0.028	1.4E-07	91,409	0.004	5.4E-01	63,601	2.7E-03
rs6971365	yes		KLF14	7	130083021	C/T	30%	0.027	2.8E-08	116,043	-0.006	2.6E-01	92,416	2.9E-06
rs9991328			FAM13A	4	89932144	T/C	49%	0.027	1.2E-09	111,934	0.007	1.4E-01	92,564	1.7E-03
rs7917772			SFXN2	10	104477433	A/G	62%	0.027	6.2E-09	113,982	0.001	8.0E-01	90,756	1.3E-04
rs2956993	yes		GANAB	11	62162738	G/T	38%	0.026	1.9E-08	111,837	0.004	4.2E-01	90,047	1.2E-03
rs8066985	yes		KCNJ2	17	65964940	A/G	51%	0.026	5.4E-09	114,268	0.005	3.5E-01	93,518	8.0E-04
rs17185536	yes		SIM1	6	100727652	C/T	76%	0.024	4.3E-05	88,603	-0.017	1.9E-02	62,861	5.5E-06
rs9648211	yes		ISPD	7	16056277	A/G	57%	0.023	3.6E-06	93,196	-0.011	6.9E-02	67,611	6.5E-06
rs3805389			NIMU	4	56177507	A/G	28%	0.023	7.1E-06	110,897	-0.013	1.9E-02	88,609	1.1E-06
rs3088050	yes		NSD1	5	176659241	A/G	21%	0.010	7.8E-02	112,933	0.036	3.0E-09	91,432	1.1E-03
rs6088735			EDEM2	20	33209337	C/T	77%	0.009	1.0E-01	114,266	0.035	6.9E-10	90,782	4.0E-04
rs7307410	yes		LEMD3	12	63828845	C/G	26%	-0.005	3.6E-01	89,227	0.033	5.0E-07	65,085	7.5E-06
rs6088552	yes		PIGU	20	32690152	G/A	37%	-0.007	1.0E-01	116,320	0.022	8.7E-06	92,396	7.2E-06
rs972303	yes		CDH10	5	24391312	T/C	75%	-0.008	1.6E-01	87,302	0.032	1.9E-06	64,371	4.1E-06
rs4898764	yes		GPNAT1	14	52334821	G/A	53%	-0.013	2.6E-03	114,264	0.016	8.5E-04	90,762	4.2E-06
rs17470444	yes		SGCZ	8	14852373	A/G	71%	-0.014	1.4E-02	86,472	0.029	1.0E-05	61,247	4.0E-07
rs2069664	yes		IQGAP2	5	75952190	G/A	53%	-0.015	3.3E-03	88,448	0.019	9.1E-04	65,083	5.7E-06
rs741361	yes		SLC2A3	12	7966952	A/G	60%	-0.016	1.7E-03	89,766	0.022	2.7E-04	64,771	8.6E-07
rs2673140	yes		IRS1	2	226868111	G/A	38%	-0.017	2.0E-04	114,393	0.018	4.1E-04	92,271	1.7E-07

(Continued)

Table 2. (Continued)

SNP	NovelLocus <sup>a</sup>	Novel Sexdiff <sup>b</sup>	NearestGene	Chr	Pos	Alleles <sup>c</sup> EA/OA	EAF	Women			Men		
								$\beta$	P	N	$\beta$	P	N
rs2042995	yes	yes	TTN	2	179266611	C/T	23%	-0.018	2.3E-03	66,222	0.022	1.4E-03	91,408
rs10881574	yes	yes	RXRA	9	136345043	C/T	7%	-0.032	2.2E-03	88,999	0.045	4.0E-04	62,482
rs7042428	yes	yes	PTPRD	9	8252414	A/G	98%	-0.053	4.3E-03	99,878	0.073	2.9E-04	79,410
rs17809093	yes	yes	CEOR2	22	16370258	G/C	4%	-0.062	8.7E-04	67,968	0.071	6.5E-04	47,894

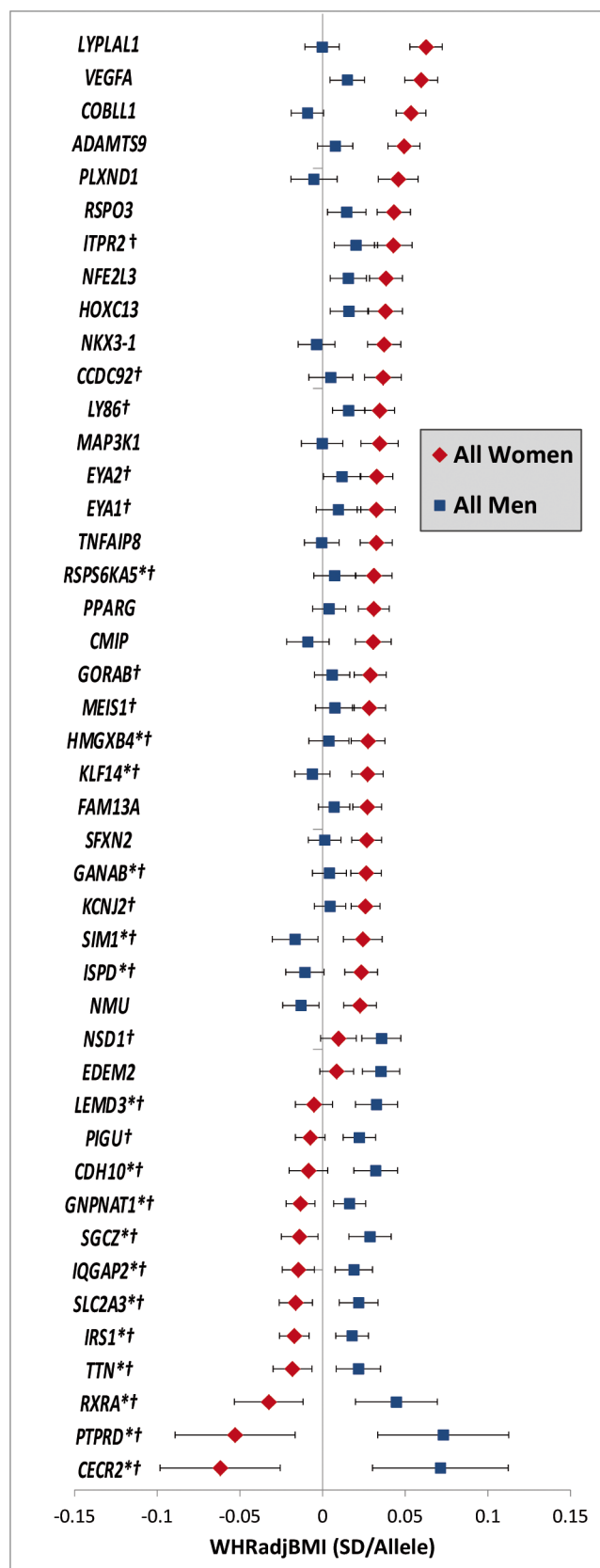
Chr: Chromosome; Pos: position; EAF: Effect Allele Frequency; EA: Effect allele; OA: Other allele

<sup>a</sup> 'Yes' if the locus is mentioned as WHR<sub>adjBMI</sub> locus for the first time

<sup>b</sup> 'Yes' if the sex-difference in the effect on WHR<sub>adjBMI</sub> is reported for the first time

<sup>c</sup> Effect allele is according to the WHR<sub>adjBMI</sub> increasing allele according to the associated sex.

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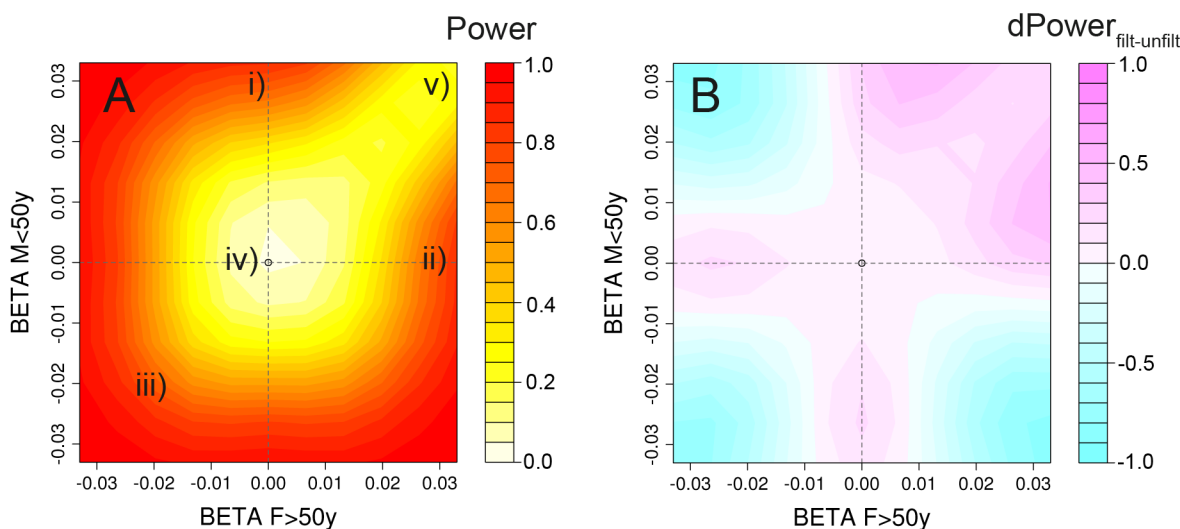


**Fig 3. Sex-dependent  $WHR_{adjBMI}$  loci.** Effect estimates ( $\beta \pm 95CI$ ) per standard deviation in  $WHR_{adjBMI}$  and risk allele for loci showing sex-differences in women compared to men. Loci are ordered by greater magnitude of effect in women compared to men. (95%CI: 95% confidence interval; SD: standard deviation. \*Newly identified loci. † Newly identified sex-differences)

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effects in blood pressure) [30], Global Lipids Genetics Consortium (GLGC) (sex-specific effects in lipids) [31], DIAGRAM (sex-specific effects for type 2 diabetes) [32] and MAGIC (sex-specific effects of glycemic traits, personal communication) [33] (S6–S10 Tables). Only CHARGE, Global-BPGen and ICBP had previously performed GWAS searching for age-specific effects on blood pressure [34]. None of the 15 age-specific BMI-associated loci influenced blood pressure in an age-specific manner ( $P_{SNP \times AGE} > 0.0033 = 0.05/15$ ) (S6 Table). Eight of the 44 sexually dimorphic  $WHR_{adjBMI}$  loci show directionally consistent female-specific effects in other traits (S10 Table), but none attained significant sex-difference ( $P_{sexdiff} > 0.0011 = 0.05/44$ ).

In addition, we performed a systematic search in the National Human Genome Research Institute (NHGRI) GWAS Catalog ([www.genome.gov/gwastudies](http://www.genome.gov/gwastudies)) to examine previously reported GWAS-associations for potential age- or sex-specificity for the loci we identified for BMI and  $WHR_{adjBMI}$ , respectively [35]. While no associations have been reported that corroborate the sex- or age-specificity of our findings, largely because few sex-stratified and no age-stratified genome-wide studies have been performed to date (this study is among the first ones), many main-effect associations with a wide range of traits and disease have been reported for our age- or sex-specific BMI or  $WHR_{adjBMI}$  loci (S11 and S12 Tables). For example, the four loci that showed a larger effect in the older group are known for their association with type 2 diabetes (T2D, near *TCF7L2* and *COBLL1*) or with coronary artery disease (CAD, near *SLC22A3* and *APOC1*). The fact that disease status may correlate both with age and obesity traits may confound our age- or sex-specific findings. To reduce this possibility we repeated



**Fig 4. Power heatplots.** Power for the combination of screens and gain through a priori filtering for varying configurations of effect sizes across the 4 strata. The figures illustrate (A) the power to detect age-difference, sex-difference or age-sex-difference in at least one of our scans (on  $P_{agediff}$ ,  $P_{sexdiff}$  and  $P_{agesexdiff}$ , with and without a priori filtering); and (B) a power comparison, comparing approaches with and without a priori filtering on  $P_{Overall} < 1 \times 10^{-5}$ . We here assume four equally sized strata and a total sample size of  $N = 300,000$  (comparable to the sample size in our BMI analyses). We set  $b_{F \leq 50y} = 0.033$  (corresponding to a known and mean BMI effect in *MAP2K5* region with  $R^2 = 0.037\%$ ),  $b_{M > 50y} = 0$ , and vary  $b_{F > 50y}$  and  $b_{M \leq 50y}$  on the axes. This strategy allows us to cover the most interesting and plausible interaction effects: Two-way interactions, such as (i) pure age-difference ( $b_{\leq 50y} = 0.033$ ,  $b_{> 50y} = 0$ ) and (ii) pure sex-difference ( $b_F = 0.033$ ,  $b_M = 0$ ); and three-way interactions, such as (iii) extreme three-way interaction with opposite direction across AGE and SEX, (iv) 1-strata interaction ( $b_{F \leq 50y} = 0.033$ ,  $b_{F > 50y} = b_{M \leq 50y} = b_{M > 50y} = 0$ ), and (v) 3-strata interaction ( $b_{F \leq 50y} = b_{F > 50y} = b_{M \leq 50y} = 0.033$ ,  $b_{M > 50y} = 0$ ).

doi:10.1371/journal.pgen.1005378.g004

the meta-analyses restricted to population-based samples (excluding all case-control studies) and observed similar effect sizes compared to the original meta-analysis (S13 and S14 Tables).

## Age-specific effects of BMI loci extend across the life course

We then examined whether the age-specific effects of the 15 BMI loci extend to younger ages and across the life course by performing look-ups in (i) a GWAS for birth weight [36] and for childhood obesity [37] from the Early Growth Genetics (EGG) Consortium, (ii) a GWAS for BMI of individuals aged 16–25 years [38], and (iii) a GWAS for weight change during adulthood (personal communication).

We found no evidence of association with *birth weight* (N = 26,836) for any of our 15 age-dependent BMI-associated loci (S15 Table) [36]. In contrast, we observed nominal significant associations with risk of *childhood obesity* (N = 13,648) for 10 of the 11 variants with stronger effect on BMI in the younger adults (Tables 3 and S16). The four loci that only showed association with BMI in the older adults were not associated with childhood obesity risk (S16 Table) [37].

Furthermore, nine of the 11 variants with stronger effect on BMI in the younger adults (18–50y) showed directionally consistent association with increased BMI in the youngest 16–25y age-group (N = 29,880, Tables 3 and S17). A more detailed experimental examination of effect sizes across the three age-groups did not reveal significant trends (S12 Fig, S17 Table, and S1 Text).

Finally, we speculated that a higher genetic BMI effect in the younger adults would translate into weight loss and a higher genetic BMI effect in the older adults would translate into weight gain with increasing age (Methods). Five of the 15 loci with age-specific effects on BMI showed a nominal significant association accompanied by the hypothesized direction on *weight change* (N = 39,041, Tables 3 and S18).

In summary, the age-dependency of the 15 loci is supported by directionally consistent enrichment of nominal significant associations ( $P < 0.05$ ) with *childhood obesity*, with BMI in the 16–25y age-group and with *weight changes* across adulthood ( $P_{\text{Binomial}}$  ranging from  $2.4 \times 10^{-5}$  to  $1.0 \times 10^{-15}$ , Table 3).

**Table 3. Enrichment analyses using look-up data for the 15 age-group specific BMI loci.** The look-up data is taken from the EGG consortium for birth weight and for childhood obesity, and from personal communication for weight change trajectories. More details including SNP specific effect sizes or odds ratios and association P-Values on the look-up trait can be found in S15 Table (for birth weight), S16 Table (for childhood obesity) and S18 Table (for weight change).

Look-up data set	Sample size	#SNPs tested	#SNPs concordant with the $\leq 50y$ vs $>50y$ association pattern	$P_{\text{binomial}}^a$	Loci with expected association pattern
Birth weight	26,836	11	0 <sup>b</sup>	$>0.99$	-
Childhood obesity	13,648	11	10 <sup>b</sup>	$1.0 \times 10^{-15}$	FTO, TMEM18, MC4R, ADCY3, NEGR1, TNNT3K, SEC16B, CBLN4, DDC, STK33
16–25y age-group	29,880	11	9 <sup>b</sup>	$2.0 \times 10^{-13}$	FTO, TMEM18, MC4R, ADCY3, NEGR1, TNNT3K, SEC16B, CBLN4, Intergenic
Weight change	39,041	15	5 <sup>c</sup>	$2.4 \times 10^{-5}$	FTO, STK33, TCF7L2, SLC22A3, APOC1

<sup>a</sup> One-sided binomial P-values that test for enrichment of nominal significant and directionally consistent association in the look-up data.

<sup>b</sup> For the BMI increasing alleles of the 11 SNPs with stronger effect on BMI in  $\leq 50y$ , we expect to see a nominal significant association with increased birth weight, increased risk for childhood obesity and increased BMI in the 16–25y age-group.

<sup>c</sup> For the BMI increasing alleles of the 11 SNPs with stronger effect on BMI in  $\leq 50y$ , we expect to see a nominal significant association with negative effect on weight change (weight loss), and for the BMI increasing alleles of the four SNPs with stronger effect on BMI in  $>50y$ , we expect to see a nominal significant association with positive effect on weight change (weight gain) (see Methods for details).

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## eQTL analysis

**eQTLs in humans.** We performed sex-specific *cis* eQTL analyses in lymphoblastoid cell lines of the combined Groningen and EGCUT studies (1,450 men and 910 women) [39, 40] for the 44 SNPs showing sex-specific effects for  $WHR_{adjBMI}$  to determine whether there is evidence to support sex-specific regulatory effects of the index variants on adjacent gene expression. Two SNP-gene associations displayed significant differences in genetic effects on expression between men and women ( $FDR(P_{Sexdiff}) < 5\%$  with and without initial filtering on overall expression effects): rs6088552–*ACSS2* and rs6088735–*MYH7B* (S19 Table). While both SNPs were associated with  $WHR_{adjBMI}$  in men-only (and no effect in women), the first SNP showed no effect on gene expression in men but was associated with gene expression in women, and the second SNP rs6088735 was associated with gene expression in both sexes, but higher in men and lower in women. The two loci were located at only 519kb from each other (rs6088552 near *PIGU*, rs6088735 near *EDEM2*, at chr20:33–34Mb,  $r^2 = 0.07$ ), each showing independent sex-specific associations with  $WHR_{adjBMI}$  and each also showing independent sex-specific association with the expression of two different genes (*ACSS2* and *MYH7B*, respectively) (S13 Fig). *ACSS2* (acyl-CoA synthetase short-chain family member 2) is a cytosolic enzyme, transcribed by SREB-proteins, that catalyzes the production of acetyl-CoA for use in both lipid synthesis and energy generation acids [41]. *MYH7B* (myosin, heavy chain 7B, cardiac muscle, beta) encodes a heavy chain subunit for slow-twitch myosin, largely expressed in heart and skeletal muscle tissue, and is involved in ATP-hydrolysis.

Age-stratified analysis were not performed for EGCUT as the study participants were relatively young (mean age: 37y), with too few individuals in the >50y age-group. Instead, we examined association between the 15 age-specific loci and gene expression using data from 3,489 unrelated individuals ( $N = 2,531$  for <50y,  $N = 958$  for ≥50y) from the NESDA and NTR cohorts [42, 43]. No SNP showed a significant age-specific effect on gene expression ( $FDR(P_{agediff}) > 5\%$  for all SNP-gene expression combinations).

**eQTLs in mice.** We compared expression of genes harboured by the identified loci in inguinal and gonadal fat in age-matched male, female or ovariectomized female (OVX) C57/BL6 mice maintained on a high-fat (HF) diet [44].

For genes located in the 15 age-specific BMI-associated loci, we compared expression in OVX female mice with the expression in the other male and female mice, but no differences in gene expression were observed.

For genes located in the 44 sex-specific  $WHR_{adjBMI}$ -associated loci, we compared expression in female mice (OVX and non-OVX) with the expression in male mice. The expression of two genes reached significance ( $P < 6.4 \times 10^{-4} = 0.05/(39 \times 2)$ ), corrected for testing 39 genes with homologous regions, and two tissues). The expression of *IQGAP2*, which regulates cell adhesion and motility, (rs2069664) was higher ( $P = 2.3 \times 10^{-7}$ ) in gonadal fat tissue of male compared to female mice, whereas the expression of *TP53INP2*, a co-factor for the thyroid hormone receptor, (rs6088552) was higher ( $P = 2.3 \times 10^{-6}$ ) in inguinal fat tissue of male compared to female mice. *TP53INP2* is located in the same chromosomal region for which we found evidence for sex-specific associations with the expression of *ACSS2* and *MYH7B* in humans. Interestingly, *Tp53inp2* has also been named the DOR (Diabetes and Obesity Related) gene, as its expression is substantially reduced in skeletal muscle of obese diabetic fa/fa Zucker rats [45]. Muscle-specific overexpression of *Tp53inp2* in mice leads to reduced muscle mass, whereas a deletion leads to muscle hypertrophy [46]. *TP53INP2* expression was markedly reduced in muscle from individuals with type 2 diabetes and in rodent diabetes models [46].



## Pathway analyses

We applied pathway analyses to gain insight into mechanisms that might be involved in the age- and sex-specific difference in body size and body shape. We assumed that loci even with moderate evidence for age- or sex-difference for BMI and  $\text{WHR}_{\text{adjBMI}}$ , respectively, are enriched for genes that contribute to the age-specific BMI association or sex-specific  $\text{WHR}_{\text{adjBMI}}$  association (**Methods**). We used the DEPICT software to perform gene set enrichment and gene expression analyses [47] (**S20** and **S21 Tables** and **S1 Text**), and QIAGEN's Ingenuity Pathway Analysis (IPA, QIAGEN Redwood City, [www.qiagen.com/ingenuity](http://www.qiagen.com/ingenuity)) tool for pathway analysis and functional annotation (**S22–S26 Tables** and **S1 Text**). Both the DEPICT and the IPA analyses identify the possible influence of sex-specific  $\text{WHR}_{\text{adjBMI}}$  loci in androgen biosynthesis, a hormone known to decrease the storage of lipids in adipose tissue [48]. Additionally, PPAR $\alpha$ /RXR $\alpha$  activation, the most significant canonical pathway for loci with a greater effect on  $\text{WHR}_{\text{adjBMI}}$  in women, may be inhibited in the presence of estrogen, thus decreasing the breakdown of lipids through competitive receptor binding [49]. To fully understand the possible age- and sex- specific regulatory effects these identified genes may have in the identified pathways, gene sets, and biological functions, further analyses are needed.

## Heritability and explained variance analyses

To assess whether the age-group differences observed for BMI and the sex-differences observed for  $\text{WHR}_{\text{adjBMI}}$  extend to the contribution of all 2.5M variants (narrow-sense heritability), we calculated heritability using the GCTA method [50] in several large studies ( $N = \text{up to } 29,232$  individuals) for all, for women and men, for the younger and older adult groups. The variance explained by the 2.5M variants was 21% for BMI and 10% for  $\text{WHR}_{\text{adjBMI}}$ , with no significant difference between age groups for BMI ( $P_{\text{agediff}} = 0.19$ ) or between men and women for  $\text{WHR}_{\text{adjBMI}}$  ( $P_{\text{sexdiff}} = 0.48$ ) (**S27 Table**).

To further investigate differences between subgroups, we calculated the variance explained in the discovery data set for subsets of SNPs based on varying thresholds of overall association on BMI or  $\text{WHR}_{\text{adjBMI}}$  (**S14 Fig**). When we included only SNPs that reached genome-wide significance for BMI ( $P_{\text{Overall}} < 5 \times 10^{-8}$ ), the variance explained in the younger adults (3.4%) was significantly larger than in the older (2.45%) adults. As we increased the significance threshold and included more SNPs with less significant overall association, the difference between the two age groups reduced and became non-significant once SNPs with a  $P_{\text{Overall}} > 3 \times 10^{-5}$  were included. We observed similar significant differences in explained variance for  $\text{WHR}_{\text{adjBMI}}$  between men and women, with the most pronounced difference for genome-wide significant SNPs ( $P_{\text{Overall}} < 5 \times 10^{-8}$ , women 1.60%; men: 0.70%) that reduced and became non-significant for SNPs with a  $P_{\text{Overall}} > 1 \times 10^{-5}$ . Consistent with the observed interactions, we found no difference in explained variance between men and women for BMI or between the younger and the older group for  $\text{WHR}_{\text{adjBMI}}$  at any  $P_{\text{Overall}}$  cut-off (**S14 Fig**).

Family-based heritability estimates, from the Family Heart Study ( $N = 1,810$ , 454 families), showed similar (but non-significant) trends for younger versus older adults for BMI (60% vs 45%,  $P_{\text{agediff}} = 0.24$ ), for women and men for  $\text{WHR}_{\text{adjBMI}}$  (43% vs 38%,  $P_{\text{sexdiff}} = 0.68$ ) (**S27 Table**).

Collectively, these observations are consistent with the results of our genome-wide search, showing that genetic variants contribute more to BMI variation in younger than in older adults and more to  $\text{WHR}_{\text{adjBMI}}$  variation in women than in men. These differences are most pronounced when we test genome-wide significant SNPs only, while differences are minimized as more SNPs with weaker associations are included.

## Joint testing of main- and interaction effects yield novel loci for BMI and $WHR_{adjBMI}$

Our stratified analysis approach also offered an opportunity for discovery of novel variants influencing BMI and  $WHR_{adjBMI}$  by (i) using a joint 4df test of the main SNP effect in the presence of interaction [27] and (ii) by overall meta-analysis of the 4 strata. Both approaches increase statistical power to detect a main effect if there is evidence of heterogeneity across the strata. Of the 164 loci that reached genome-wide significance for BMI ( $P < 5 \times 10^{-8}$ ), 73 are novel (S28 Table and S1, S2 and S15 Figs). Of the 73 loci, 45 were only identified in the overall test and 26 were identified in both tests. The remaining two loci were only identified in the joint test and either displayed evidence for difference between men and women (near *CXXC5*,  $P_{sexdiff} = 2.7 \times 10^{-5}$ ) or between age-groups (near *DDC*,  $P_{agediff} = 6.2 \times 10^{-4}$ ) suggesting that its identification may have been aided by allowing for interaction. We identified 53 loci with significant associations with  $WHR_{adjBMI}$ , of which 10 were novel (S29 Table and S1, S2 and S16 Figs). It can be speculated that the yield of novel SNP associations for BMI was greater than that of  $WHR_{adjBMI}$ , because age-dependent effects have not been sought systematically before, whereas sex-specific screens have been performed previously [10].

## Discussion

Our genome-wide search for age- and sex-specific loci in up to 320,485 adults of European ancestry identified 15 loci that were associated with BMI in an age-dependent manner, with predominantly larger effects in the younger than in the older adults. Notably, despite sufficient statistical power, we did not identify BMI-associated loci with sex-dependent effects. The largest association study on BMI [19] identified two SNPs with different impact on BMI in men and women: rs543874 (*SEC16B*) and rs6091540 (*ZFP64*). While these SNPs show more modest trends towards sex-different effect ( $P_{sexdiff} = 2.4 \times 10^{-4}$  and  $1.3 \times 10^{-4}$ , respectively) in our study, they were not picked up by our analysis due to the different pre-filtering strategy. In contrast to BMI and consistent with previous observations for  $WHR_{adjBMI}$ , we identified 44  $WHR_{adjBMI}$  associated loci with sex-specific effects of which the majority have a larger effect in women compared with men. No age-specific  $WHR_{adjBMI}$  loci were discovered.

Our work is the first large-scale genome-wide association study to interrogate the influence of both age and sex, simultaneously, on genetic effects for BMI and  $WHR_{adjBMI}$ . While our meta-analysis had sufficient power to identify SNP-by-age or SNP-by-sex interactions, we only discovered loci influenced by age for BMI. Studies that followed up on previously established BMI loci in longitudinal and cross-sectional designs support our findings regarding the age-dependency of the majority of these loci [38, 51–57]. Indeed, for 11 of the 15 loci identified in our study, the effect on BMI was 1.5 to 3.5 times smaller in the older adults than in the younger adults, which may reflect a greater culmination of environmental and lifestyle factors on adiposity in older adults that overwhelm the genetic effects. While none of these loci were associated with birth weight, all—but one—were nominally associated with increased risk of childhood obesity. Results from a GWAS on BMI in 16-to-25 year-olds [58] provide preliminary evidence that some loci exert their largest effects relatively early in life, whereas others become more pronounced in young adulthood. Notwithstanding the predominance of BMI loci with larger genetic effects in younger individuals we identified four loci with stronger genetic effects in older adults. Interestingly, these four loci have been previously associated with either type 2 diabetes [32] or coronary artery disease [59]. Sensitivity analyses precluded potential ascertainment bias introduced by disease studies in the older group. These loci may influence BMI through mechanisms that are distinct from other BMI-associated loci; mechanisms that may be more closely related to processes more directly involved in the pathogenesis

obesity-related diseases. Furthermore, the directional consistent genetic effects of our loci on weight change during adult life from longitudinal studies supports our finding.

Indeed, the stratification into age-groups may introduce a cohort effect that implies a different genetic or environmental make-up of cohorts with older vs younger adults. For example, the obesogenic environment that has fueled the obesity epidemic that westernized societies have experienced during the past 30 years may have affected older individuals differently than younger individuals. To examine the contribution of such cohort effects and to obtain more accurate age-dependent effect estimates, large-scale genetic longitudinal studies would be required that measure BMI at multiple time points with individuals born across a wide range of birth years.

While our study provides some first insights into age-dependent genetic effects, in particular before and after menopause, more data from larger studies with longitudinal data spanning from childhood through late adulthood are desirable to accurately assess the influence of these loci on BMI across the life course. Indeed, identifying the time of life when variants affect body weight the most may help us determine the mechanisms of their influence on body weight and potential for intervention.

In contrast to the observations for BMI, our genome-wide interaction analyses did not identify loci with age-dependent effects for  $\text{WHR}_{\text{adjBMI}}$  but there was strong novel evidence for sex-influenced effects in 44 loci. For 27 of the 44 loci, the sexual dimorphism is reported for the first time, with 17 being completely novel associations for  $\text{WHR}_{\text{adjBMI}}$ . Due to increased sample size and optimized SNP selection approaches, we more than doubled the number of loci with established sex-difference for  $\text{WHR}_{\text{adjBMI}}$  [10, 18, 29]. The 44 loci divide into 11 loci with opposite effects between men and women, 28 loci with a stronger effect in women and five loci with a stronger effect in men. This is the first report to highlight loci with opposite effects and the enrichment of women-specific  $\text{WHR}_{\text{adjBMI}}$  associations is consistent with previous findings.

We examined whether the sex-dependent effects on  $\text{WHR}_{\text{adjBMI}}$  were mediated through sex-specific effects on the expression of genes located within these loci, using data available from eQTL analyses in humans and mice. Of particular interest is a region at chromosome 20q11.22 in which two independent  $\text{WHR}_{\text{adjBMI}}$  lead SNPs near *PIGU* and near *EDEM2* showed independent sex-specific associations with the expression of *ACSS2* and *MYH7B*, respectively, in humans. While we found no direct evidence of sex-specific action of *ACSS2* or *MYH7B*, based on current knowledge, both proteins seem to be involved in peripheral energy metabolism. In addition, we observed that the expression of *Tp53inp2* (Tumor Protein 53 Inducible Nuclear Protein 2), of which the human *TP53INP2* ortholog is also located in the *PIGU* locus, had significantly higher expression levels in the inguinal fat of male than female mice. This observation is consistent with a previous study, showing that *Tp53inp2* expression in white adipose tissue is significantly higher in male than in female mice [60]. The authors speculated that this sex-specificity might be due to differences in fat distribution with females storing proportionally more fat in subcutaneous/inguinal and males more in intra-abdominal depots [60]. Taken together, the sex-specific association with  $\text{WHR}_{\text{adjBMI}}$  of two independent loci at chr20q11.22 may be mediated through any or all three genes for which we found sex-specific expression. While all three genes are good candidates, experimental follow up will be needed to pinpoint the causal gene(s) and to elucidate the function and sex-specificity.

Our broad-sense (family-based analyses) or narrow-sense (GCTA including all 2.5M variants) heritability estimates showed no difference in explained variance between men or women, or between younger and older adults for either outcome. However, when considering subsets of variants displaying overall significant associations ( $P_{\text{Overall}} < 1 \times 10^{-5}$ ), we observed a significant difference between age- but not sex- groups for BMI, with a larger explained

variance among the younger than the older adults, and between sex- but not age groups for  $WHR_{adjBMI}$ , with a larger explained variance in women than in men. These observations further corroborate the predominance of age-dependent loci for BMI and sex-dependent loci for  $WHR_{adjBMI}$  identified through a genome-wide screen.

Even though our study is likely the largest GxE and the first  $G \times E_1 \times E_2$  interaction GWAS meta-analysis ever conducted, we did not detect loci with sex-specific effects for BMI (SNP x SEX), age-specific effects for  $WHR_{adjBMI}$  (SNP x AGE) or three-way interactions effects (SNP x AGE x SEX). Three-way interactions are biologically plausible when considering that sex-specific effects might be exerted through hormones and that the hormonal status particularly of women changes at menopause (i.e., around the age of 50 years). This would result in a 1-stratum interaction (i.e., genetic effect only present in younger women) or a 3-strata interaction (i.e., genetic effect present in all but in younger women). While our study had sufficient power (power > 80%) to identify any kind of two-way interaction (SNP x SEX or SNP x AGE) even for effects as small as those observed for established BMI or  $WHR_{adjBMI}$  loci, our power was limited specifically for the biologically plausible three-way effects (1-stratum or 3-strata-interaction). To detect subtle effects appearing in only one of the four strata will require specialized study designs or alternative approaches. We provide a detailed analytical perspective on the power to detect different interaction signals that may inform other studies aiming at detecting interaction effects.

We acknowledge that our power estimations are expressed as a function of previously observed explained variances, incorporating measurement error. As measurement error increases, the variance of the phenotype increases and—because the genetic effect is not affected—the explained variance of the genetic variants decreases. While a random measurement error in the dependent variable of a linear regression model would not lead to a biased effect size estimate, such an error would increase the standard errors of the effect size estimates compared to a measurement error free outcome. Under the alternative hypothesis, this results in smaller statistical power. This would imply, for our analysis, that we have potentially missed some true associations, which could have been detected with smaller measurement error.

With the growing sample-size and thus statistical power, measurement error is often larger than the variant-wise effect size estimate for many human traits currently under investigation in large-scale GWAS. Thus, an individual variant's effect may not have clinical significance by itself in predictive models. However, its ultimate significance should be evaluated in the context of the biological mechanism it reveals along with other discovered variants, and the potential of such a mechanism as a therapeutic target; this is yet to be determined. In order to discover more disease-associated genetic variants, reducing measurement error by repeated and/or more accurate measurements is a viable alternative to only increasing sample size—especially when the measurement error relative to the outcome variability is high.

For technical reasons, variants on the X-chromosome were not screened. Yet, an interesting hypothesis is that sex-linked variants contribute to a sex-dependent architecture of body size and shape, both of which exhibit obvious sexual dimorphism. These analytic challenges are being addressed currently, and exploration of X-linked variation is warranted. Further, we have included only individuals of European-ancestry and thus cannot report on the generalizability of our findings to other race or ethnic groups. While we examined age-dependent effects by binning individuals below and above age 50 years—an average age of menopause—it is possible that modeling of age as a continuous trait might have had superior power. This approach poses more complex harmonization issues that should be addressed in a follow-up study. In addition, we recognize that environmental modifiers may further influence the effect of trait-related loci, and that some of the interactions we identified may be proxies for interactions with other environmental factors that are correlated with either age or sex.



In summary, our findings further distinguish the genetics of BMI from the genetics of  $\text{WHR}_{\text{adjBMI}}$ . Previously described aspects of distinction include the enrichment of neural pathways versus insulin-related pathways and sexual consistency versus sexual dimorphism, respectively [61, 62]. Our findings suggest that genetic BMI effects can change by age possibly depicting different mechanisms of genetic BMI effects that either increase or decrease during adult age. The knowledge of such mechanisms might guide the development of more effective intervention programs that are desperately sought after.

## Methods

### Anthropometric phenotypes

The anthropometric traits examined are body mass index (BMI,  $\text{kg/m}^2$ ), which is a measure of body mass and a surrogate for total body fat, and waist-to-hip-ratio adjusted for BMI ( $\text{WHR}_{\text{adjBMI}}$ ), which is a measure of body fat distribution. Traits were transformed before analyses; we first created age- (and BMI) adjusted residuals (including age and  $\text{age}^2$  into the regression for BMI, and additionally BMI for  $\text{WHR}_{\text{adjBMI}}$ ) for each of the four strata separately (men  $\leq 50\text{y}$ , men  $> 50\text{y}$ , women  $\leq 50\text{y}$ , and women  $> 50\text{y}$ ) and subsequently applied an inverse normal transformation.

### Study-specific analyses

We included up to 92 studies (totalling up to 21,989 men  $\leq 50\text{y}$ , 74,324 men  $> 50\text{y}$ , 41,386 women  $\leq 50\text{y}$ , and 88,625 women  $> 50\text{y}$ ) with genome-wide genotyping chip data using either Affymetrix or Illumina arrays. To enable meta-analyses across different SNP panels, each study group performed genotype imputation using HapMap II CEU (build 21 or 22) via MACH [63], IMPUTE [64] or BimBam [65] yielding  $\sim 2.8$  Million SNPs. In addition, we included 22 studies (up to 28,106, 18,877, 29,306, 17,872 individuals for each of the strata, respectively) for BMI and  $\text{WHR}_{\text{adjBMI}}$  that were genotyped using the custom iSELECT Metabochip array containing  $\sim 195\text{K}$  SNPs designed to support large-scale follow-up of putative associations with metabolic and cardiovascular traits [66].

In each study, SNP associations were tested separately by age-group and sex (men  $\leq 50\text{y}$ , men  $> 50\text{y}$ , women  $\leq 50\text{y}$  and women  $> 50\text{y}$ ) for autosomal variants. The additive genetic effect for each SNP on each phenotype was estimated via linear regression using MACH2QTL [67], SNPTTEST [64], ProbABEL [68], GenABEL [69], Merlin [70], PLINK [71] or QUICKTEST [72]. For studies with a case-control design, cases and controls were analysed separately. See [S1](#), [S2](#) and [S3 Tables](#) for study specific genotyping, imputation, analysis, quality control and phenotypic descriptive information. In total we gathered association data from up to 92 studies with imputed GWAS data and 22 studies genotyped on the Metabochip array for BMI including up to 320,485 individuals and 64 studies with imputed GWAS data and 20 studies genotyped on the Metabochip array for  $\text{WHR}_{\text{adjBMI}}$  including up to 216,654 individuals.

All studies were conducted according to the principles expressed in the Declaration of Helsinki. The studies were approved by the local Review Boards and all study participants provided written informed consent for the collection of samples and subsequent analysis.

### Quality control of study-specific aggregated data

All study-specific files were processed in the meta-analysis centers through a standardized quality-control (QC) pipeline [73]. This involved QC checks on file completeness, range of test statistics, allele frequencies, trait transformation and population stratification as well as filtering on low quality data. Briefly, we excluded monomorphic SNPs, SNPs with  $\text{MAF}^*N \leq 3$

(minor allele frequency multiplied by sample size), imputed SNPs with poor imputation quality:  $r^2_{\text{hat}} < 0.3$  in MACH, observed/expected dosage variance  $< 0.3$  in BAMBAM,  $\text{proper\_info} < 0.4$  in IMPUTE, information  $< 0.8$  in PLINK [64, 65, 67, 71]; genotyped SNPs with low call-rate ( $< 95\%$ ), and genotyped SNPs that were out of Hardy-Weinberg equilibrium (HWE, P-Value testing for HWE  $< 10^{-5}$ ). To increase the overlap in the number of SNPs between imputed GWAS and MetaboChip data, we transferred all SNP identifiers to unique SNP names consisting of chromosomal and base position, e.g. using chr1:217820132 instead of rs2820443 in the meta-analysis. Sex- and age-specific standard errors and P-values from each participating study were genomic-control (GC) corrected using study- and strata-specific lambda factors [74], whereas the lambdas were estimated from all genome-wide available SNPs for imputed GWAS and form a subset of 4,427 QT-interval SNPs for MetaboChip studies.

## The meta-analyses

Generally, beta-estimates and standard errors were meta-analyzed using an inverse-variance weighted fixed effect model as implemented in METAL [75].

We meta-analyzed effect estimates and standard errors from all available studies in each of the four strata separately, yielding  $b_{M \leq 50y}$ ,  $b_{M > 50y}$ ,  $b_{F \leq 50y}$ ,  $b_{F > 50y}$  and  $SE_{M \leq 50y}$ ,  $SE_{M > 50y}$ ,  $SE_{F \leq 50y}$ ,  $SE_{F > 50y}$ . By meta-analyzing  $b_{M \leq 50y}$  and  $b_{M > 50y}$  we obtained the effect and standard error for men ( $b_M$ ,  $SE_M$ ) and women ( $b_F$ ,  $SE_F$ ). Similar meta-analyses yielded the age group-specific association statistics,  $b_{\leq 50y}$  and  $b_{> 50y}$  with standard errors  $SE_{\leq 50y}$  and  $SE_{> 50y}$ . Meta-analysis of all four strata provided the overall association effect estimate  $b_{\text{overall}}$ , standard error  $SE_{\text{overall}}$  and P-value  $P_{\text{overall}}$ . A joint meta-analysis based on the pooled stratum-specific estimates was performed according to Aschard et al [27].

After the meta-analyses, we performed an additional quality control step on the meta-analytic results: We only included SNPs (i) being available in at least half of the maximum sample size in all strata; and (ii) having chromosome and position annotation in dbSNP.

## Genome-wide screening approaches to detect interaction effects

Our study aimed at discovering SNPs with (1) **age-different** effects, (2) **sex-different** effects, and (3) **age-dependent sex-different** effects or **sex-dependent age-different** effects.

To find **age-different** genetic effects, we computed age-difference P-values ( $P_{\text{agediff}}$ ) by testing for difference between the age group-specific meta-analyzed beta-estimates  $b_{\leq 50y}$  and  $b_{> 50y}$  using

$$t_{\text{age}} = \frac{b_{\leq 50y} - b_{> 50y}}{\sqrt{SE_{\leq 50y}^2 + SE_{> 50y}^2 - 2r_{\text{age}} \cdot SE_{\leq 50y} \cdot SE_{> 50y}}}.$$

The correlation  $r_{\text{age}}$  between  $b_{\leq 50y}$  and  $b_{> 50y}$  computed as the Spearman rank correlation coefficient across all SNPs for BMI and  $\text{WHR}_{\text{adjBMI}}$  was 0.123 and 0.049, respectively. The analogous test statistic for **sex-different** effects was

$$t_{\text{sex}} = \frac{b_M - b_F}{\sqrt{SE_M^2 + SE_F^2 - 2r_{\text{sex}} \cdot SE_M \cdot SE_F}},$$

with corresponding P-value ( $P_{\text{sexdiff}}$ ). The Spearman correlation  $r_{\text{sex}}$  was 0.121 or 0.047 for BMI and  $\text{WHR}_{\text{adjBMI}}$ , respectively.

To test for the three-way interaction of age- and sex-differences, we introduced for the first time a test of difference between age groups in the sex-difference, which is mathematical equivalent to a test of difference between sexes in the age group-difference using the age-sex-

difference statistic as

$$t_{agesex} = \frac{(b_{M \leq 50y} - b_{F \leq 50y}) - (b_{M > 50y} - b_{F > 50y})}{\sqrt{SE_{M \leq 50y}^2 + SE_{F \leq 50y}^2 + SE_{M > 50y}^2 + SE_{F > 50y}^2}},$$

with the corresponding P-value ( $P_{agesexdiff}$ ).

To maximize statistical power we did not split our samples (artificially) into discovery and replication sets, but meta-analyzed all studies together and verified the absence of cross-study heterogeneity. We screened genome-wide for  $P_{agediff}$ ,  $P_{sexdiff}$ , and  $P_{agesexdiff}$  for each of the two traits (BMI,  $WHR_{adjBMI}$ ). These screens have ideal power to detect effects that are of opposite direction across the four strata (S9 Fig). However, searching for effects that are prominent in one or some strata, but not existent or directionally consistent and less pronounced in other strata profits from an a priori filter on the overall association ( $P_{overall} < 10^{-5}$ ) as shown previously [10, 26] (Fig 4). The rationale behind this filter is that SNPs with unequal effects in the different strata have non-zero overall effect when tested in all strata combined. This is true unless these effects are the same magnitude, but in opposite direction (i.e. cancel out in the combined analysis). Hence filtering on overall association P-value possibly enriches our selection with SNPs showing interaction effects. For BMI and  $WHR_{adjBMI}$  7,382 and 2,014 SNPs passed this filter.

For each trait and for each of the 6 approaches ( $P_{agediff}$ ,  $P_{sexdiff}$ ,  $P_{agesexdiff}$ , with and without a priori filtering), we controlled the False Discovery Rate (FDR) at 5% to account for the multiple testing [76]. Importantly, controlling the FDR of each single analysis at 5% implies a global FDR control at 5% for the ensemble of discoveries resulting from all the different approaches together.

## Sensitivity analyses using population-based studies only

To ensure the association of none of our age- or sex-specific loci were driven by ascertainment bias through inclusion of case-series of individuals with type 2 diabetes or coronary artery disease, we performed additional meta-analyses restricted to population-based (i.e. no ascertainment bias) studies and compared the effect-sizes between the original meta-analyses and the meta-analyses restricted to population-based studies.

## Sensitivity analyses excluding studies with self-reported BMI or WHR

Self-reported BMI or WHR may cause systematic measurement error that might lead to biased effect estimates. Few of our studies assessed BMI and WHR by self-report in the sense that they told study participants how to measure BMI and WHR for themselves. In order to ensure that the age- or sex-differences of our identified loci was not driven by the few studies that used self-report data (13 of our 114 studies), which may introduce bias [77–79], we conducted sensitivity meta-analyses limited to studies that measured anthropometric phenotypes (S5 and S8 Figs).

## Power computations

To illustrate the strength and characteristics of the various screens outlined, we analytically computed power by scan (S9 Fig) and for all scans combined (Figs 4, S10 and S11), for varying configurations of effect size combinations and directions across the four strata. More specifically, we assumed equally sized strata, a total sample size approximately corresponding to the maximum sample size of our study and modelled three categories of SNPs explaining realistic fractions of the phenotypic variance, i.e. small, medium and large effects from Seliotes et al

[28] and from Heid et al [29]. The power shown in any of the heatplots was calculated based on a fixed effect in women  $\leq 50y$  (set to the known effect), a fixed effect in men  $> 50y$  (set to 0), and varying effects in women  $> 50y$  and men  $\leq 50y$  (varying from negative to positive magnitude of the known effect). This strategy allowed us to depict power for most important interaction effects (i.e. for pure sex-difference, pure age-difference, 1-strata interaction and 3-strata interaction) in a single heatplot (see legend of Fig 4).

## Genome-wide screening approaches to detect main effects accounting for interaction

To identify novel genetic association for BMI and  $WHR_{adjBMI}$ , we screened (i) the  $P_{Overall}$  gathered from a four-way meta-analysis of the stratified results and (ii) the  $P_{Joint}$  gathered from a four-way joint meta-analysis of the stratified results according to Aschard et al [27]. We used a genome-wide significance level ( $P < 5 \times 10^{-8}$ ) for both approaches to correct for the multiple testing and compared the detected regions to previously established loci using a 500kb distance criterion.

## Establishing enrichment for sex-specific or age-dependent genetic effects

For  $WHR_{adjBMI}$ , we counted among the sex-different associations (disregarding the opposite effect loci) how many were significantly stronger in men or women. To test whether the observed counts represent significant imbalances between sexes we compared them to the expected binomial distribution (with  $p = 0.5$ ). Similar exercise was done for age-specific associations for BMI.

## Lookup of age- and sex-specific associations with other phenotypes

Age-group specific association results of the identified loci were requested for blood-pressure measures (diastolic and systolic blood pressure, mean arterial pressure and pulse pressure) from the Global-BPGen consortium [30]. The provided effect size and standard error estimates for six age bins (20–29, 30–29, . . . , 70–79 years) were combined to derive SNP x AGE interaction effect sizes and P-Values (S6 Table) using meta-regression [34].

Sex-specific associations of the identified loci were requested for lipid traits (HDL-C, LDL-C, Total Cholesterol and Triglycerides) from the Global Lipids Genetics Consortium [31], for type 2 diabetes (T2D) from the DIAGRAM consortium [32], for glycemic traits (fasting insulin, fasting glucose, HOMA-B, HOMA-IR) from the Meta-Analyses of Glucose and Insulin-related traits (MAGIC) Consortium [33] (personal communication), and for blood-pressure measures (diastolic and systolic blood pressure) from the Global-BPGen consortium [30] (S7, S8 and S9 Tables). The provided men- and women-specific estimates were used to derive sex-difference P-Values.

## NHGRI GWAS catalog lookups

To further investigate the identified genetic variants in this study and to gain additional insight into their functionality and possible pleiotropic effects, we searched for previous SNP-trait associations nearby our lead SNPs. PLINK was used to find all SNPs within 500 kb of any of our lead SNPs using 1000 Genomes Project Pilot I genotype data from the CEPH (Utah residents with ancestry from northern and western Europe) population (CEU) [80, 81]. To identify previous associations, all SNPs within the specified regions were compared with the NHGRI (National Human Genome Research Institute) catalog for overlap and distances between the



two SNPs were obtained using SAS, Version 9.2 [citation info below for SAS and PLINK] [82]. The NHGRI's (National Human Genome Research Institute) GWAS catalog contains only the top 30 most significant SNP-trait associations from recent GWAS published results from studies with at least 100,000 SNPs with resulting P-values of less than  $P < 1 \times 10^{-5}$  [82]. For previous GWAS results not reported in the Catalog when accessed on 10/15/2014, additional SNP-trait associations were pulled from the literature and compared to our lead SNPs using the same PLINK output file to obtain distance and  $r^2$  values [83–91]. All previous associations within 500 kb and with an  $r^2 > 0.1$  with our lead SNP that reached genome-wide significance in the previous publication were retained for further interrogation.

## Association of age-specific BMI loci with birth weight and childhood obesity

Summary statistics from a genome-wide association meta-analyses previously performed by EGG Consortium ([www.egg-consortium.org](http://www.egg-consortium.org)) were used to examine whether the 15 age-specific BMI loci associate with birth weight and/or childhood obesity risk. Birth weight (BW) had been transformed to z-scores. Association between each SNP and the birth weight was tested using linear regression assuming an additive genetic model, with sex and, where available, gestational age as covariables [36]. In the genome-wide association meta-analysis for childhood obesity risk, cases were defined as having an age- and sex-specific BMI > 95th percentile, and controls as having an age- and sex-specific BMI < 50th percentile in children of European ancestry. SNP associations were assessed in a case-control design assuming an additive genetic model [37].

## Comparison of effect sizes for age-dependent BMI loci with younger individuals aged 16–25 years

We compared the effect sizes for 15 loci with age related differences in BMI for each of the age strata ( $\leq 50$ y and  $> 50$ y) in men and women combined with the BMI in young adults ages 16–25 years [58]. Nine out of the 14 studies included in the young adult analysis had overlapping samples with the current sample, although the BMI measurements utilized were different (i.e. adolescence/early adulthood versus middle-aged to older adulthood). We used t-tests to compare effect estimates ( $\beta$ ) from the younger adults aged 16–25 years (A) to each of our age strata ( $\leq 50$ y or  $> 50$ y) (B) adjusting for the correlation due to overlapping samples such that:

$$t_{diff} = \frac{b_A - b_B}{\sqrt{SE_A^2 + SE_B^2 - 2r \cdot SE_A \cdot SE_B}},$$

where SE = standard error and  $r$  = Spearman correlation coefficient between the effect estimates genome-wide. We calculated the Spearman correlation  $r$  between our study and GIANT using the combined stages from both studies. The significance level (P-value) was based on a two-tailed t-test.

## Look-up of age-dependent BMI loci for weight change across adulthood

We also evaluated the 15 BMI loci showing age-dependent results from genome-wide analyses with weight change across adulthood. Using growth curves generated from multiple measures of weight in individuals between the ages of 20 and 65 years, weight change trajectories were calculated by sex using age as both a random and fixed effect. For each of the 15 loci showing age-differences in BMI, we observationally compared the direction of the effect estimate in the weight change results with the direction of effect seen between our adults aged 18–50 years and

adults >50 years. While assuming constant height across adulthood and no cohort effect between the two age-groups, we hypothesized that for loci where we find a stronger effect for BMI in the adults ages 18–50 years compared to adults >50 years, the direction of effect estimate in the weight change data would be negative. For the loci where we found a stronger effect for BMI in the adults >50 years compared to the adults ages 18–50 years we hypothesized that the direction of effect estimate in the weight change data would be positive.

## Expression QTL analyses in human tissue

We examined transcript expression of genes nearby ( $\pm 1$  Mb) the 44 identified  $WHR_{adjBMI}$  SNP in lymphoblastoid human cell lines available in 2,360 human samples from the EGCUT and Groningen cohorts (910 women and 1,450 men) [39, 40]. We computed sex-specific associations between each of the 44 variants and all genes in their 1Mb vicinity and tested the men- and women-specific eQTLs for sex-difference ( $FDR_{sexdiff} < 5\%$  calculated with/without initial filter on overall expression effect  $FDR_{overall} < 20\%$ ).

We next examined whether the 15 SNPs identified to be age-dependently associated with BMI impact nearby ( $\pm 1$  Mb) transcripts differently in younger (<50y) than in older individuals ( $\geq 50y$ ). As such, we analyzed human whole blood transcription in 3,489 unrelated individuals from NESDA and NTR cohorts [42, 43], which were divided in a  $\geq 50y$  group ( $N = 958$ ) and a <50y group ( $N = 2,531$ ). Cis-eQTL analysis for the 15 SNPs was conducted for the two groups separately and age-group specific eQTLs were compared for age-difference ( $FDR_{agediff} < 5\%$  calculated with/without initial filter on overall expression effect  $FDR_{overall} < 20\%$ ).

## Expression QTL analyses of adipose tissues in high-fat-diet-induced obese mice

We performed a microarray analysis on data from an experiment previously published [44]. Briefly, 21 male, 21 female, and 21 ovariectomized (OVX) female C57/BL6 mice were fed from day 21 for 12 weeks on a high fat diet (45% calories from fat; Research Diets, Inc., New Brunswick, NJ). All mice (male, female, and OVX) were exposed to sham or OVX surgery. Animals were sacrificed and tissues collected during the first 2h of the beginning of the light cycle after a 12h fast.

GeneChip microarray (Affymetrix, Santa Clara, CA) was performed according to manufacturer's instructions on 7 independent pooled samples (3 mice per pooled sample) per experimental group (male, female, OVX) from gonadal adipose tissue (GWAT) and inguinal adipose tissue (IWAT) fat pads. AMC Project Report Version 12 (6/27/07) GeneChip Operating System parameters  $\alpha 1$  and  $\alpha 2$  were set to 0.05 and 0.065, respectively. Normalized expression values from the Affymetrix identifier were analyzed with the online software server Genesifter (VizX Labs, Inc., Seattle, WA, USA). For comparisons of microarray data sets, multiple t-tests were used to identify genes with at least a twofold difference in gene expression (with Benjamini and Hochberg correction;  $P < 0.05$ ) and at least an expression level of 100. Genes populated from the GWAS studies were compared to this list of genes that met the minimum criteria of expression, fold difference, and p-value. Those identified as being statistically significant were further validated by qPCR.

## Pathway analyses

**DEPICT.** We used a recently developed pathway enrichment method, DEPICT [47]. The methodology first selects all lead SNPs below a certain threshold with respect to a target P-value (available genome-wide). We tested multiple hypotheses corresponding to different

lead SNP selection scenarios. First, we selected SNPs with  $P_{\text{sexdiff}} < 0.001$ . Second, focusing on SNPs with concordant effect size direction (CED), but different magnitude we added a marginal filter to boost power by selecting SNPs with  $P_{\text{sexdiff}} < 0.01$  and  $P_{\text{overall}} < 0.01$ . In case of CED, SNPs with stronger effect in women may fall into separate pathways from SNPs with stronger effect in men. Hence, we have derived gender-specific sex-difference P-values ( $P_{\text{sexdiff\_F}}$ ,  $P_{\text{sexdiff\_M}}$ ). We then looked for women-specific pathway enrichment by selecting SNPs with  $P_{\text{sexdiff\_F}} < 0.01$  and  $P_{\text{overall}} < 0.01$  (given the CED framework). Similarly, we created a separate list for men-specific SNPs by a filter of  $P_{\text{sexdiff\_M}} < 0.01$  and  $P_{\text{overall}} < 0.01$ . All above lists were also created for age-dependent BMI associations by replacing  $P_{\text{sexdiff}}$  by  $P_{\text{agediff}}$ ,  $P_{\text{sexdiff\_F}}$  by  $P_{\text{agediff\_younger}}$  and  $P_{\text{sexdiff\_M}}$  by  $P_{\text{agediff\_older}}$ .

For each of the eight SNP lists, lead SNPs were identified. For each lead SNP locus a target region is defined as the smallest interval containing all SNPs with  $LD > 0.5$  with the lead SNP of the locus. All genes encompassed in the target regions represent the “GWAS genes”, thereby assuming that either the lead SNP is in LD with a functional coding SNP within a gene or that the lead SNP marks a cis-acting regulatory region. We then used the following pre-defined gene sets and pathways: Gene Ontology (GO), Reactome, InWeb protein complexes, Mouse knock-out phenotypes. Gene sets were re-annotated based on co-expression in a large collection (80,000) of gene expression compendium from GEO. Then, for each gene-set the pairwise similarity between GWAS genes was calculated and compared to that of matching sets of non-GWAS genes to assess significance of enrichment.

DEPICT also generated a prioritized set of genes at each locus. Briefly, genes within associated loci that are functionally similar to genes from other associated loci are the more likely candidates. DEPICT prioritizes genes in three steps: gene scoring, bias adjustment, and false discovery rate estimation. In the scoring step, the method quantifies the similarity of a given gene to genes from other associated loci. The bias adjustment step controls confounding factors that may bias the gene scores, e.g. gene length. In the last step, experiment-wide false discovery rates are estimated by repeating the scoring and bias adjustment steps 20 times based on top SNPs from pre-computed null GWAS.

**Ingenuity Pathway Analysis (IPA).** Significantly associated loci were further explored using QIAGEN’s Ingenuity Pathway Analysis (IPA, QIAGEN Redwood City, [www.qiagen.com/ingenuity](http://www.qiagen.com/ingenuity)) to determine if there was an over-arching functional or disease relationship among these loci and their associated genes using the age and sex specific SNP lists described above. IPA uses publicly available databases (e.g. NHGRI GWAS Catalog, NCBI databases, KEGG) and proprietary databases of gene/protein interaction, expression, and function to identify possible pathways, networks, and overlapping functions of genes. For our analysis, IPA identified potential genes as those genes with coding regions within 2kb upstream or 0.5kb downstream of our list of input dbSNP ids that can unambiguously be mapped to these ids. To perform the analyses, only Ingenuity Knowledge Base genes were used, both direct and indirect relationships that are observed or predicted in mammals (humans, mice, and rats) are strictly considered. All canonical pathways and functional/disease categories and annotations that were statistically significant ( $P < 0.05$  using the Fisher’s exact test) are reported; however, those that meet significance for multiple test correction (Benjamin-Hochberg corrected  $P < 0.05$ ) are highlighted in the table. Only the top ten predicted networks containing up to 140 genes or endogenous molecules were requested. Only those networks with a score of greater than 2 (Fisher’s Exact Test result of  $P < 0.01$ ) are considered significant [92].

## Estimation of heritability

We estimated the broad heritability ( $H^2$ ) of BMI and  $WHR_{adjBMI}$  within the Family Heart Study (FHS) to assess how much of each trait's total phenotypic variance may be genetic. A random sample of 1,810 individuals (454 families) was used for this analysis. The sample was stratified by age and sex into 9 groups (all, all  $\leq 50y$ , all  $> 50y$ , men, women, men  $\leq 50y$ , men  $> 50y$ , women  $\leq 50y$ , women  $> 50y$ ) to assess how each trait's genetic variance may differ across strata. Within each group, BMI and  $WHR_{adjBMI}$  were adjusted for age, age<sup>2</sup>, genotyping chips (Illumina 560K, 1,000,000K, 610K), 10 principal components and 3 study centers. Residuals for BMI and  $WHR_{adjBMI}$  were ranked and an inverse normal transformation was applied. Subsequently, SOLAR was used to estimate the  $H^2$  of BMI and  $WHR_{adjBMI}$  within each group ([S28 Table](#)).

## Genome-wide Complex Trait Analysis for proportion of variance explained

To explore the contribution of all common (genotyped) SNPs genome-wide to each trait of interest, BMI and  $WHR_{adjBMI}$ , we estimated the variance explained by all the autosomal SNPs in the combined ARIC, KORA S3/S4, CoLaus, EGCUT and SHIP studies within each of the sex and age strata, using the method proposed by Yang et al [93] and implemented in the Genome-wide Complex Trait Analysis software package (GCTA <http://www.complextaitgenomics.com/software/gcta/>). Each phenotypic trait was transformed in the same form as was used for all meta-analyses.

## Estimation of explained variance

We estimated the age-group and sex-specific polygenic variance explained by various subsets of SNPs that were based on varying thresholds of overall association ( $P_{Overall}$ ) with BMI or  $WHR_{adjBMI}$ . First, each subset of SNPs was clumped into independent regions using a physical distance criterion  $< 500kb$  and for each region the most significantly overall associated SNP (i.e. top SNP) was taken further. For each top SNP, the explained variance was calculated according to

$$r^2 = \frac{1}{1 + \frac{N}{(\Phi^{-1}(\frac{p}{2}))^2}} - \frac{1 - r^2}{N}$$

for each age-group and for each sex separately [94]. Finally, the variance explained by the subset of SNPs was obtained by summing up the single SNP-specific explained variances. The overall association threshold was varied from  $1 \times 10^{-8}$  to 0.1.

## Search for biological and functional knowledge of the identified association regions

We examined whether SNPs known to provide reliable tags for Copy-Number-Variations (CNVs) in subjects of European-descent (combining four catalogues including 60,167 CNV-tagging SNPs as described previously [95]) correlated with our lead SNPs. We also performed several online database searches to establish whether known variants within a 500kb-window on both sides of each lead SNP, that are in high linkage disequilibrium ( $r^2 > 0.8$ ) with our lead SNPs (using SNAP Proxy search [96]), might have putative or predicted function. (i) We searched the SIFT database [97] to determine whether any of these SNPs were predicted to affect protein function. (ii) We used Annovar [98] to investigate predicted and putative



function in several functional classes, including splicing regulation, stop codons, polyphen predictions. (iii) We used the regulome database (<http://regulome.stanford.edu/>) to search for known and predicted regulatory elements (DNAase hypersensitivity, binding sites of transcription factors, and promoter regions) in the intergenic regions of our age-specific BMI and sex-specific  $WHR_{adjBMI}$  loci. Additionally, we searched for estrogen, androgen or progesterone receptor motifs around our sex-specific  $WHR_{adjBMI}$  loci. Source of these data include public datasets from GEO, the ENCODE project, and published literature [99].

## Supporting Information

**S1 Fig. Workflow and overview of results.** The numbers stated are the number of identified independent loci for the respective analysis. Given in brackets is the number of the identified loci that are novel loci for the trait, i.e. have not been previously reported for association with the trait.

(TIF)

**S2 Fig. QQ plots for overall association and joint test P-Values for both traits.** QQ-plots for BMI (A) and  $WHR_{adjBMI}$  (B) depicting overall association P-Values (red) and joint test P-Values (blue) for all SNPs and after excluding previously published BMI or  $WHR_{adjBMI}$  associated regions ( $P_{Overall}$ : magenta;  $P_{Joint}$ : cyan).

(TIF)

**S3 Fig. Locuszoom plots for 15 loci associated with BMI that are different between men and women  $\leq 50$ y and men and women  $> 50$ y.** Each plot highlights the most significant SNP for age-differences and illustrates p-values for age-differences ( $P_{agediff}$ ), sex-differences ( $P_{sexdiff}$ ), all strata combined ( $P_{Overall}$ ), and the joint test ( $P_{Joint}$ ). The figure is sorted according to Table 1. The plots are based on GrCh37 build positions and annotations.

(TIF)

**S4 Fig. Scatterplot of effect estimates (beta) for loci showing age-differences in BMI, contrasting loci with larger effect estimates in men & women  $\leq 50$  years (light green diamonds) and loci with larger effects in men & women  $> 50$  years (dark green squares).**

(TIF)

**S5 Fig. Sensitivity meta-analysis for the 15 age-specific BMI loci-excluding 13 studies that used self-report data for BMI and comparing the age-difference effects to the originally observed age-difference.**

(TIF)

**S6 Fig. Locuszoom plots for 44 loci associated with  $WHR_{adjBMI}$  that are different between men and women.** Each plot highlights the most significant SNP for sex-differences and illustrates p-values for age-differences ( $P_{agediff}$ ), sex-differences ( $P_{sexdiff}$ ), all strata combined ( $P_{Overall}$ ), and the joint test ( $P_{Joint}$ ). The figure is sorted according to Table 2. The plots are based on GrCh37 build positions and annotations.

(TIF)

**S7 Fig. Scatterplot of effect estimates (beta) for loci showing sex-differences in waist-to-hip ratio adjusted for BMI ( $WHR_{adjBMI}$ ), organized by loci with larger effect estimates in women compared to men (red circles), larger effect estimates in men compared to women (blue squares) and opposite effect estimates between men and women (green triangles).**

(TIF)

**S8 Fig. Sensitivity meta-analysis for the 44 sex-differential  $WHR_{adjBMI}$  loci—excluding two self-report studies and comparing the sex-difference effects to the originally observed sex-difference.**

(TIF)

**S9 Fig. Power by AGE x SEX scan.** The figures illustrate the power of scanning  $P_{sexdiff}$  (A: unfiltered, B: pre-filtered on  $P_{Overall}$ ),  $P_{agediff}$  (C: unfiltered, D: pre-filtered on  $P_{Overall}$ ), and  $P_{agesexdiff}$  (E: unfiltered, F: pre-filtered on  $P_{sexdiff}$  or on  $P_{agediff}$ ). We assume four equally sized strata, a total sample size of  $N = 300,000$  (comparable to the sample size in our BMI analyses). To investigate varying scenarios of interaction effects, we set (i)  $b_{F<50y} = 0.033$ , a median BMI effect near *MAP2K5* from Speliotes et al. ( $R^2 = 0.037\%$ ), (ii)  $b_{M>50y} = 0$ , and (iii) vary  $b_{F>50y}$  and  $b_{M<50y}$  on the x- and y-axes respectively.

(TIF)

**S10 Fig. Power of the AGE x SEX approaches for BMI for varying allele frequencies and varying modelled effect sizes.** The figure shows the power to detect age-difference, sex-difference or age x sex-difference in at least one of our scans and for varying scenarios of effect size combinations between the 4 strata. We assume four equally sized strata and a total sample size of  $N = 300,000$  (comparable to the sample size in our BMI analyses). Furthermore, for each plot we (i) set  $b_{F<50y}$  to a known BMI effect sizes from Speliotes et al. paper (using a small (*PTPB2*), medium (*NEGR1*) and the largest (*FTO*) effect size), (ii) set  $b_{M>50y} = 0$ , and (iii) vary  $b_{F>50y}$  and  $b_{M<50y}$  on the axes.

(TIF)

**S11 Fig. Power of the AGE x SEX approaches for  $WHR_{adjBMI}$  for varying allele frequencies and varying modelled effect sizes.** The figure shows the power to detect age-difference, sex-difference or age x sex-difference in at least one of our scans and for varying scenarios of effect size combinations between the 4 strata. We assume four equally sized strata and a total sample size of  $N = 200,000$  (comparable to the sample size in our  $WHR_{adjBMI}$  analyses). Furthermore, for each plot we (i) set  $b_{F<50y}$  to a known  $WHR_{adjBMI}$  effect sizes from Heid et al. paper (using a small (*CPEB4*), medium (*LYPLAL1*) and the largest (*RSPO3*) effect size), (ii) set  $b_{M>50y} = 0$ , and (iii) vary  $b_{F>50y}$  and  $b_{M<50y}$  on the axes.

(TIF)

**S12 Fig. Differences in effect estimates ( $\beta \pm SE$ ) between young adults, adults  $\leq 50y$ , and adults  $> 50y$  for BMI loci selected for age-differences.** Loci are ordered according to trends in absolute magnitude of effect: 1) where the absolute magnitude of effect is largest in adolescent/youngest adults (ages 16–25y)<sup>1</sup>, 2) where absolute magnitude is largest in adults ( $\leq 50y$ ), and 3) where absolute magnitude is largest in older adults ( $> 50y$ ). BMI: Body mass index; SE: standard error; Details for men and women ages 16–25 have been described elsewhere (Graff et al.: “Genome-wide analysis of BMI in adolescents and young adults reveals additional insight into the effects of genetic loci over the life course.” Human Molecular Genetics 2013).

(TIF)

**S13 Fig. The most significant SNPs, rs6088552 and rs6088735, for sex-differences with  $WHR_{adjBMI}$  each identified to be a sex-different cis-eQTL for the *ACSS2* and *MYH7B* genes, respectively, on chromosome 20.**  $WHR_{adjBMI}$ : waist-to-hip ratio adjusted for body-mass index; eQTL: expression quantitative trait loci. Sex-specific associations were computed to identify cis eQTL signals that were likely to be coincident with the  $WHR_{adjBMI}$  using human eQTL in lymphoblastoid cells.

(TIF)

**S14 Fig. Total stratum-specific explained variance by SNPs meeting varying thresholds of overall association for BMI (A: sex-specific; B: age-group specific) and for  $WHR_{adjBMI}$  (C: sex-specific; D: age-specific).**

(TIF)

**S15 Fig. Locuszoom plots for 73 novel loci associated with BMI that were either identified by the joint 4df test or by the overall (age-group and sex—combined) analysis.** Each plot highlights the most significant SNP for the combined effect ( $P_{Overall}$ ) or for the joint test ( $P_{Joint}$ ) and illustrates p-values for age-differences ( $P_{Agediff}$ ), sex-differences ( $P_{Sexdiff}$ ) and  $P_{Joint}$  or  $P_{Overall}$  respectively<sup>a</sup>. The figure is sorted according to chromosome and position. The plots are based on GrCh37 build positions and annotations. For three loci we identified two different SNPs that met the significance threshold for the scan of  $P_{Overall}$  and  $P_{Joint}$ . For each set we plotted the SNP with the lowest P-value based on the scan it was identified for. These loci and the SNP plotted are as follows: 1) rs7421089 – Selected for  $P_{Joint}$  and rs10804189 – Selected for  $P_{Overall}$ ->rs10804189 is plotted, 2) rs1557765 – Selected for  $P_{Overall}$  and rs7928810 – Selected for  $P_{Joint}$ -> rs7928810 is plotted, and 3) rs11181001 – Selected for  $P_{Joint}$  & rs1405552 – Selected for  $P_{Overall}$ -> rs1405552 is plotted.

(TIF)

**S16 Fig. Locuszoom plots for 10 novel loci associated with  $WHR_{adjBMI}$  that were either identified by the joint 4df test or by the overall (sex-combined) analysis.** Each plot highlights the most significant SNP for the combined effect ( $P_{Overall}$ ) or for the joint test ( $P_{Joint}$ ) and illustrates p-values for age-differences ( $P_{Agediff}$ ), sex-differences ( $P_{Sexdiff}$ ) and  $P_{Joint}$  or  $P_{Overall}$  respectively. The figure is sorted according to chromosome and position. The plots are based on GrCh37 build positions and annotations.

(TIF)

**S1 Table. Study design, number of individuals and sample quality control for genome-wide association study cohorts.**

(XLSX)

**S2 Table. Information on genotyping methods, quality control of SNPs, imputation, and statistical analysis for genome-wide association study cohorts.**

(XLSX)

**S3 Table. Study-specific descriptive statistics of study cohorts.** \*\* There were significant differences in the number of subjects available for different phenotypes. In this case, separate summary statistics were provided.

(XLSX)

**S4 Table. Stratum-specific results and extended details for the 15 age-specific BMI loci.** The table is ordered according to [Table 1](#).

(XLSX)

**S5 Table. Stratum-specific results and extended details for the 44 sex-specific  $WHR_{adjBMI}$  loci.** The table is ordered according to [Table 2](#).

(XLSX)

**S6 Table. Age-specific associations of age-dependent BMI SNPs with blood pressure (BP).** Abbreviations: Effect Allele (EA), Other Allele (OA), SNP-by-age interaction effect ( $b_{SNP \times AGE}$ ), SNP-by-age interaction effect standard error ( $SE_{SNP \times AGE}$ ), SNP-by-age interaction P-value ( $P_{SNP \times AGE}$ ).

(XLSX)

**S7 Table. Sex-specific associations of sexually dimorphic  $WHR_{adjBMI}$  SNPs with lipid traits (GLGC).** Abbreviations: Effect Allele (EA), Other Allele (OA), High Density Lipoprotein Cholesterol (HDL), Low Density Lipoprotein (LDL), Total Cholesterol (TC), Triglycerides (TG). (XLSX)

**S8 Table. Sex-specific associations of sexually dimorphic  $WHR_{adjBMI}$  SNPs with Type 2 Diabetes (T2D, DIAGRAM) and glycemic traits (MAGIC).** Abbreviations: Effect Allele (EA), Other Allele (OA), Odds Ratio (OR). (XLSX)

**S9 Table. Sex-specific associations of sexually dimorphic  $WHR_{adjBMI}$  SNPs with blood pressure (BP) measures.** Abbreviations: Effect Allele (EA), Other Allele (OA), Odds Ratio (OR). (XLSX)

**S10 Table. Remarkable women-specific associations in the  $WHR_{adjBMI}$  lookup data.** The table shows SNPs that meet a Bonferroni-corrected significance level ( $<0.05/44$ ) for its sex-specific association with the lookup trait and no association with the lookup trait in the other sex. Only women-specific loci displayed similar patterns in the look-up data. None of the opposite effect direction loci showed opposite effects (requesting  $P < 0.05$  in both sexes) with a look-up trait. (XLSX)

**S11 Table. Previously-identified associations listed in the NHGRI GWAS Catalog that lie within 500 kb and  $r^2 > 0.1$  to our lead BMI SNPs.** (XLSX)

**S12 Table. Previously-identified associations listed in the NHGRI GWAS Catalog that lie within 500 kb and  $r^2 > 0.1$  to our lead  $WHR_{adjBMI}$  SNPs.** (XLSX)

**S13 Table. BMI loci showing significant age-differences in adults  $\leq 50y$  compared to adults  $>50y$ .** Analysis was restricted to non-case control studies. (XLSX)

**S14 Table.  $WHR_{adjBMI}$  loci showing significant sex-differences.** Analysis was restricted to non-case control studies. (XLSX)

**S15 Table. Effect estimates of BMI loci selected for age-differences and birthweight from 26,836 participants in the EGG consortium.** (XLSX)

**S16 Table. Odds ratios of BMI loci selected for age-differences and childhood obesity from 5,530 cases and 8,318 controls in the EGG consortium.** (XLSX)

**S17 Table. Differences in effect estimates between young adults, adults  $\leq 50y$ , and adults  $>50y$  for BMI loci selected for age-differences.** (XLSX)

**S18 Table. Effect estimates for weight change trajectories in adults between the ages of 20 and 65 years of age in loci showing effect size differences in BMI by age.** (XLSX)



**S19 Table. Top hits of the human sex-specific eQTL lookup of the sex-specific  $WHR_{adjBMI}$  associated SNPs.** Presented are SNPs with significant sex-differences in eQTL effects, selected according to  $FDR(P\text{-Sexdiff}) < 5\%$ , with and without initial filtering on overall expression effects ( $FDR(P\text{-Overall}) < 20\%$ ).

(XLSX)

**S20 Table. List of tissues in which DEPICT identified significant expression ( $FDR < 0.1$ ) of genes from age-specific BMI associated loci in at least one of the four approaches.**

(XLSX)

**S21 Table. Gene sets enriched ( $FDR < 0.1$ ) for harboring SNPs with sex-different effect on  $WHR_{adjBMI}$  identified by DEPICT.**

(XLSX)

**S22 Table. All significant function or disease annotations identified in IPA for the younger adult-specific BMI loci.** Functions that remain significant after B-H ( $p < 0.05$ ) correction are marked in bold.

(XLSX)

**S23 Table. All significant function or disease annotations identified in IPA for the older adult-specific BMI loci.** Functions that remain significant after B-H correction are marked in bold.

(XLSX)

**S24 Table. All significant canonical pathways identified in IPA for the women-specific  $WHR_{adjBMI}$  loci.** Pathways that remain significant after B-H correction are marked in bold.

(XLSX)

**S25 Table. All significant function or disease annotations identified in IPA for the women-specific  $WHR_{adjBMI}$  loci.** Functions that remain significant after B-H correction are marked in bold.

(XLSX)

**S26 Table. All significant function or disease annotations identified in IPA for the men-specific  $WHR_{adjBMI}$  loci.** Functions that remain significant after B-H correction are marked in bold.

(XLSX)

**S27 Table. Proportion of genetic to phenotypic variance explained for BMI, and  $WHR_{adjBMI}$  estimated using GCTA and Heritability estimated using SOLAR.**

(XLSX)

**S28 Table. BMI main or joint (main+interaction, 4df) effect findings compared to results from the GIANT BMI group [19].**

(XLSX)

**S29 Table.  $WHR_{adjBMI}$  main or joint (main+interaction, 4df) effect findings compared to results from the GIANT WAIST group [18].**

(XLSX)

**S1 Text. Supplementary note.**

(DOCX)

**S2 Text. Consortia members and extended acknowledgments.**

(DOCX)

## Author Contributions

Conceived and designed the experiments: IBB IMH KEN RJFL TWW ZK. Performed the experiments: AEJ LBa MGr TWW ZK. Analyzed the data: AEJ LBa MGr RM TWW ZK. Contributed reagents/materials/analysis tools: AEJ LBa MGr TWW ZK. Wrote the paper: AEJ IBB IMH KEN LBa MGr MFF RJFL TWW ZK. Contributed genome-wide association study results: AAH AB ACA ACHa ACHe ACS AD AEF AEJ AFW AG AGU AHa AHo AJ ÅJ AJMdc AJO AJS AKH ALJ ALMV AMa AMe APa APe APM APo ARSa ARSh ASB ASG ASst ATe ATH ATö ATr AUJ AVS AWM BAO BDM BF BL BMK BMP BOB BSt BT BWP CAB CAH CBa CBe CBo CG CHa CHe CLang CM CMG CML CMvD CO CR CSF CW CWKC DA DAB DAE DC DCR DFL DH DIB DIC DJH DPS DS EA EB EE EI EJCdG EK EM EPB ETs EV EW EZ FB FC FF FG FH FLM FRe FRiv FRiz FSC FWA GB GC GDed GDel GE GL GM GP GRA GT GWa GWi GWM HAB HAK HC HGra HGrö HH HMDr HMS HScha HSchu HSsn HV HVH HWal HWat IB IBB IF IK IMH IML IMN IN IPi IPr IR IT JAS JBe JBl JCB JCC JCM JCo JCT JcZ JD JDR JEH JErd JEri JFW JG JGE JH JHS JHZ JIR JJH JKr JKu JLa JLBG JLu JMJ JMV JNH JPT JROC JSB JSh JSin JSK JSP JSV JT JvS JvVO JWJ JY KAR KBJ KEN KKO KLMoh KLMon KMR KP KSL KSte KSti KTK LAC LALMK LBa LBr LCG LCPGMdG LFe LJL LJP LK LLa LLi LLW LM LMR LMYA LPas LPat LPé LQ LS LV LYe LYu MASa MASw MBa MBl MBo MBr MCV MCZ MD MEK MEM MF MFF MGo MGP MGr MHa MHe MJ MKä MKaa MKar MLa MLL MLo MMa MMe MN MP MRh MS MSN MWald MWalk ND NE NF NGI NGM NH NHK NJW NLHC NLP NMvS NP NS NV NvdV NWR OG OPo OTR PAFM PB PBK PD PEHS PES PF PGH PH PIWdB Pjvdm PK PKM PLdJ PMR PPP PS PSC PV PvdH PVG PWF RAS RJFL RL RM RNB RPS RR RTG RWW SA SB SCvD SE SG SH SHV SHW SIB SJ SJC SJK SJLB SKan SKat SKo SL SMä SRB SRW SSa SSc SSeb ST STT SWvdL TAL TBG TBH TDS TE TEG TFa TH TIAS TK TKR TL TLA TMT TQ TR TSA TT TW UG UT UV VG VLo VLy VSa VSt VV WLMA WM WRS WZ YCC YDIC YJS YLi YLu YMG YPL ZK ZW. Contributed Metabochip association study results: AB AHa AHi AL AMe ASi AW BG BOB BSe CLang CLanz DB DK EAA EE EI EM ETr FB FRe IB IMH JHZ JLi JLu JSa KB KF KGH KH KKO KL KSti KTK LLB LLH LM LYe MGo MKi MKu MMN MRi MU NGF NGr NJW NLP NN NTK OH OPe PD PF PM PWF RAS RJFL RJS RL RRM RW SA SCB SKan SL SMe TE TFa TH THS TI TJ TS TSA TWW Udf Wko WKr. Contributed SNP look-up data: AC CCW CEJ DCR GBE IPr JSim LAC NLHC RM SSen TFe VLa. Performed Study-specific GCTA analyses: ATe MGr SR TE TWW ZK. Performed study-specific quality control: AEJ JcZ LBa MFF MGr RM SC TE TFa TOK TWW YLu ZK. Performed DEPICT Gene Pathway analyses: JKa JNH LFr THP. Performed IPA Gene Pathway analyses: AEJ. Performed Expression Quantitative trait loci (eQTL) analyses: HJW LFr RJ TE. Performed Mouse microarray expression analyses: DJC MDN. Contributed Look-up information and analyses for young adults ages 16–25 years: KEN LAC MGr PGL SIB. GIANT Steering committee: CMvD CSF DJH DPS DS EI GRA IB IBB IMH JNH JROC KEN KLMoh LCG MBo MIMC PD RCK RJFL SIB TLA UT.

## References

1. Vazquez G., et al., Comparison of body mass index, waist circumference, and waist/hip ratio in predicting incident diabetes: a meta-analysis. *Epidemiol Rev*, 2007. 29: p. 115–28. PMID: [17494056](#)
2. Pischon T., et al., General and abdominal adiposity and risk of death in Europe. *N Engl J Med*, 2008. 359(20): p. 2105–20. doi: [10.1056/NEJMoa0801891](#) PMID: [19005195](#)
3. Mokdad A.H., et al., Prevalence of obesity, diabetes, and obesity-related health risk factors, 2001. *JAMA*, 2003. 289(1): p. 76–9. PMID: [12503980](#)
4. Must A., et al., The disease burden associated with overweight and obesity. *JAMA*, 1999. 282(16): p. 1523–9. PMID: [10546691](#)

5. Yusuf S., et al., Obesity and the risk of myocardial infarction in 27,000 participants from 52 countries: a case-control study. *Lancet*, 2005. 366(9497): p. 1640–9. PMID: [16271645](#)
6. Canoy D., et al., Body fat distribution and risk of coronary heart disease in men and women in the European Prospective Investigation Into Cancer and Nutrition in Norfolk cohort: a population-based prospective study. *Circulation*, 2007. 116(25): p. 2933–43. PMID: [18071080](#)
7. De Mello J.J., et al., Gender Differences In The Evaluation Of Adult Body Composition. *Medicine and Science in Sports and Exercise*, 2005. 37: p. S299–S299.
8. Kirchengast S., Gender Differences in Body Composition from Childhood to Old Age: An Evolutionary Point of View. *Journal of Life Sciences*, 2010. 2(1): p. 1–10.
9. Legato M.J., Beyond women's health the new discipline of gender-specific medicine. *Med Clin North Am*, 2003. 87(5): p. 917–37, vii. PMID: [14621324](#)
10. Randall J.C., et al., Sex-stratified Genome-wide Association Studies Including 270,000 Individuals Show Sexual Dimorphism in Genetic Loci for Anthropometric Traits. *Plos Genetics*, 2013. 9(6): p. e1003500. doi: [10.1371/journal.pgen.1003500](#) PMID: [23754948](#)
11. Walter A.A., et al., Sarcopenia Indices: Age- And Gender-related Differences In Body Composition, Strength, And Muscle Quality. *Medicine and Science in Sports and Exercise*, 2012. 44: p. 98–98.
12. Wells J.C., Sexual dimorphism of body composition. *Best Pract Res Clin Endocrinol Metab*, 2007. 21(3): p. 415–30. PMID: [17875489](#)
13. Lombar-Albrecht L.A. and Styne D.M., Effect of puberty on body composition. *Curr Opin Endocrinol Diabetes Obes*, 2009. 16(1): p. 10–5. PMID: [19115520](#)
14. Rogol A.D., Roemmich J.N., and Clark P.A., Growth at puberty. *J Adolesc Health*, 2002. 31(6 Suppl): p. 192–200. PMID: [12470915](#)
15. Rosenbaum M. and Leibel R.L., Clinical review 107: Role of gonadal steroids in the sexual dimorphisms in body composition and circulating concentrations of leptin. *J Clin Endocrinol Metab*, 1999. 84(6): p. 1784–9. PMID: [10372664](#)
16. Kuk J.L., et al., Age-related changes in total and regional fat distribution. *Ageing Res Rev*, 2009. 8(4): p. 339–48. doi: [10.1016/j.arr.2009.06.001](#) PMID: [19576300](#)
17. Mott J.W., et al., Relation between body fat and age in 4 ethnic groups. *American Journal of Clinical Nutrition*, 1999. 69(5): p. 1007–1013. PMID: [10232643](#)
18. Shungin D., et al., New genetic loci link adipose and insulin biology to body fat distribution. *Nature*, 2015. 518(7538): p. 187–96. doi: [10.1038/nature14132](#) PMID: [25673412](#)
19. Locke A.E., et al., Genetic studies of body mass index yield new insights for obesity biology. *Nature*, 2015. 518(7538): p. 197–206. doi: [10.1038/nature14177](#) PMID: [25673413](#)
20. Abdunour J., et al., The effect of the menopausal transition on body composition and cardiometabolic risk factors: a Montreal-Ottawa New Emerging Team group study. *Menopause*, 2012. 19(7): p. 760–7. doi: [10.1097/gme.0b013e318240f6f3](#) PMID: [22395454](#)
21. Douchi T., et al., Precedence of bone loss over changes in body composition and body fat distribution within a few years after menopause. *Maturitas*, 2003. 46(2): p. 133–138. PMID: [14559384](#)
22. Morita Y., et al., Precedence of the shift of body-fat distribution over the change in body composition after menopause. *Journal of Obstetrics and Gynaecology Research*, 2006. 32(5): p. 513–516. PMID: [16984520](#)
23. Bromberger J.T., et al., Prospective study of the determinants of age at menopause. *Am J Epidemiol*, 1997. 145(2): p. 124–33. PMID: [9006309](#)
24. Gold E.B., et al., Factors associated with age at natural menopause in a multiethnic sample of midlife women. *Am J Epidemiol*, 2001. 153(9): p. 865–74. PMID: [11323317](#)
25. Gold E.B., et al., Factors Related to Age at Natural Menopause: Longitudinal Analyses From SWAN. *Am J Epidemiol*, 2013. 178(1): p. 70–83. doi: [10.1093/aje/kws421](#) PMID: [23788671](#)
26. Kooperberg C. and Leblanc M., Increasing the power of identifying gene x gene interactions in genome-wide association studies. *Genet Epidemiol*, 2008. 32(3): p. 255–63. doi: [10.1002/gepi.20300](#) PMID: [18200600](#)
27. Aschard H., et al., Genome-wide meta-analysis of joint tests for genetic and gene-environment interaction effects. *Hum Hered*, 2010. 70(4): p. 292–300. doi: [10.1159/000323318](#) PMID: [21293137](#)
28. Speliotes E.K., et al., Association analyses of 249,796 individuals reveal 18 new loci associated with body mass index. *Nat Genet*, 2010. 42(11): p. 937–48. doi: [10.1038/ng.686](#) PMID: [20935630](#)
29. Heid I.M., et al., Meta-analysis identifies 13 new loci associated with waist-hip ratio and reveals sexual dimorphism in the genetic basis of fat distribution. *Nature Genetics*, 2010. 42(11): p. 949–60. doi: [10.1038/ng.685](#) PMID: [20935629](#)

30. Newton-Cheh C., et al., Genome-wide association study identifies eight loci associated with blood pressure. *Nat Genet*, 2009. 41(6): p. 666–76. doi: [10.1038/ng.361](https://doi.org/10.1038/ng.361) PMID: [19430483](https://pubmed.ncbi.nlm.nih.gov/19430483/)
31. Teslovich T.M., et al., Biological, clinical and population relevance of 95 loci for blood lipids. *Nature*, 2010. 466(7307): p. 707–13. doi: [10.1038/nature09270](https://doi.org/10.1038/nature09270) PMID: [20686565](https://pubmed.ncbi.nlm.nih.gov/20686565/)
32. Morris A.P., et al., Large-scale association analysis provides insights into the genetic architecture and pathophysiology of type 2 diabetes. *Nat Genet*, 2012. 44(9): p. 981–90. doi: [10.1038/ng.2383](https://doi.org/10.1038/ng.2383) PMID: [22885922](https://pubmed.ncbi.nlm.nih.gov/22885922/)
33. Scott R.A., et al., Large-scale association analyses identify new loci influencing glycemic traits and provide insight into the underlying biological pathways. *Nat Genet*, 2012. 44(9): p. 991–1005. doi: [10.1038/ng.2385](https://doi.org/10.1038/ng.2385) PMID: [22885924](https://pubmed.ncbi.nlm.nih.gov/22885924/)
34. Simino J., et al., Gene-Age Interactions in Blood Pressure Regulation: A Large-Scale Investigation with the CHARGE, Global BPgen, and ICBP Consortia. *Am J Hum Genet*, 2014. 95(1): p. 24–38. doi: [10.1016/j.ajhg.2014.05.010](https://doi.org/10.1016/j.ajhg.2014.05.010) PMID: [24954895](https://pubmed.ncbi.nlm.nih.gov/24954895/)
35. Hindroff, L.A., et al., *A Catalog of Published Genome-Wide Association Studies Available at [www.genome.gov/gwastudies](http://www.genome.gov/gwastudies)*. 2010.
36. Horikoshi M., et al., New loci associated with birth weight identify genetic links between intrauterine growth and adult height and metabolism. *Nat Genet*, 2013. 45(1): p. 76–82. doi: [10.1038/ng.2477](https://doi.org/10.1038/ng.2477) PMID: [23202124](https://pubmed.ncbi.nlm.nih.gov/23202124/)
37. Bradfield J.P., et al., A genome-wide association meta-analysis identifies new childhood obesity loci. *Nat Genet*, 2012. 44(5): p. 526–31. doi: [10.1038/ng.2247](https://doi.org/10.1038/ng.2247) PMID: [22484627](https://pubmed.ncbi.nlm.nih.gov/22484627/)
38. Graff M., et al., The influence of obesity-related single nucleotide polymorphisms on BMI across the life course: the PAGE study. *Diabetes*, 2013. 62(5): p. 1763–7. doi: [10.2337/db12-0863](https://doi.org/10.2337/db12-0863) PMID: [23300277](https://pubmed.ncbi.nlm.nih.gov/23300277/)
39. Leitsalu L., et al., Cohort Profile: Estonian Biobank of the Estonian Genome Center, University of Tartu. *Int J Epidemiol*, 2014.
40. Fehrmann R.S., et al., Trans-eQTLs reveal that independent genetic variants associated with a complex phenotype converge on intermediate genes, with a major role for the HLA. *PLoS Genet*, 2011. 7(8): p. e1002197. doi: [10.1371/journal.pgen.1002197](https://doi.org/10.1371/journal.pgen.1002197) PMID: [21829388](https://pubmed.ncbi.nlm.nih.gov/21829388/)
41. Luong A., et al., Molecular characterization of human acetyl-CoA synthetase, an enzyme regulated by sterol regulatory element-binding proteins. *J Biol Chem*, 2000. 275(34): p. 26458–66. PMID: [10843999](https://pubmed.ncbi.nlm.nih.gov/10843999/)
42. Wright F.A., et al., Heritability and genomics of gene expression in peripheral blood. *Nat Genet*, 2014. 46(5): p. 430–7. doi: [10.1038/ng.2951](https://doi.org/10.1038/ng.2951) PMID: [24728292](https://pubmed.ncbi.nlm.nih.gov/24728292/)
43. Jansen R., et al., Sex differences in the human peripheral blood transcriptome. *BMC Genomics*, 2014. 15: p. 33. doi: [10.1186/1471-2164-15-33](https://doi.org/10.1186/1471-2164-15-33) PMID: [24438232](https://pubmed.ncbi.nlm.nih.gov/24438232/)
44. Grove K.L., et al., A microarray analysis of sexual dimorphism of adipose tissues in high-fat-diet-induced obese mice. *Int J Obes (Lond)*, 2010. 34(6): p. 989–1000.
45. Baumgartner B.G., et al., Identification of a novel modulator of thyroid hormone receptor-mediated action. *PLoS One*, 2007. 2(11): p. e1183. PMID: [18030323](https://pubmed.ncbi.nlm.nih.gov/18030323/)
46. Sala D., et al., Autophagy-regulating TP53INP2 mediates muscle wasting and is repressed in diabetes. *J Clin Invest*, 2014. 124(5): p. 1914–27. doi: [10.1172/JCI72327](https://doi.org/10.1172/JCI72327) PMID: [24713655](https://pubmed.ncbi.nlm.nih.gov/24713655/)
47. Pers T.H., et al., Biological interpretation of genome-wide association studies using predicted gene functions. *Nat Commun*, 2015. 6: p. 5890. doi: [10.1038/ncomms6890](https://doi.org/10.1038/ncomms6890) PMID: [25597830](https://pubmed.ncbi.nlm.nih.gov/25597830/)
48. Veilleux A., et al., Glucocorticoid-induced androgen inactivation by aldo-keto reductase 1C2 promotes adipogenesis in human preadipocytes. *Am J Physiol Endocrinol Metab*, 2012. 302(8): p. E941–9. doi: [10.1152/ajpendo.00069.2011](https://doi.org/10.1152/ajpendo.00069.2011) PMID: [22275760](https://pubmed.ncbi.nlm.nih.gov/22275760/)
49. Yoon M., The role of PPARalpha in lipid metabolism and obesity: focusing on the effects of estrogen on PPARalpha actions. *Pharmacol Res*, 2009. 60(3): p. 151–9. doi: [10.1016/j.phrs.2009.02.004](https://doi.org/10.1016/j.phrs.2009.02.004) PMID: [19646654](https://pubmed.ncbi.nlm.nih.gov/19646654/)
50. Yang J., et al., Common SNPs explain a large proportion of the heritability for human height. *Nat Genet*, 2010. 42(7): p. 565–9. doi: [10.1038/ng.608](https://doi.org/10.1038/ng.608) PMID: [20562875](https://pubmed.ncbi.nlm.nih.gov/20562875/)
51. Hardy R., et al., Life course variations in the associations between FTO and MC4R gene variants and body size. *Hum Mol Genet*, 2010. 19(3): p. 545–52. doi: [10.1093/hmg/ddp504](https://doi.org/10.1093/hmg/ddp504) PMID: [19880856](https://pubmed.ncbi.nlm.nih.gov/19880856/)
52. Hertel J.K., et al., FTO, type 2 diabetes, and weight gain throughout adult life: a meta-analysis of 41,504 subjects from the Scandinavian HUNT, MDC, and MPP studies. *Diabetes*, 2011. 60(5): p. 1637–44. doi: [10.2337/db10-1340](https://doi.org/10.2337/db10-1340) PMID: [21398525](https://pubmed.ncbi.nlm.nih.gov/21398525/)
53. den Hoed M., et al., Genetic susceptibility to obesity and related traits in childhood and adolescence: influence of loci identified by genome-wide association studies. *Diabetes*, 2010. 59(11): p. 2980–8. doi: [10.2337/db10-0370](https://doi.org/10.2337/db10-0370) PMID: [20724581](https://pubmed.ncbi.nlm.nih.gov/20724581/)

54. Graff M., et al., Estimation of genetic effects on BMI during adolescence in an ethnically diverse cohort: The National Longitudinal Study of Adolescent Health. *Nutr Diabetes*, 2012. 2: p. e47. doi: [10.1038/nutd.2012.20](https://doi.org/10.1038/nutd.2012.20) PMID: [23168566](https://pubmed.ncbi.nlm.nih.gov/23168566/)
55. Murphy R.A., et al., Candidate Gene Association Study of BMI-Related Loci, Weight, and Adiposity in Old Age. *J Gerontol A Biol Sci Med Sci*, 2013. 68(6): p. 661–6. doi: [10.1093/gerona/gls227](https://doi.org/10.1093/gerona/gls227) PMID: [23160366](https://pubmed.ncbi.nlm.nih.gov/23160366/)
56. Elks C.E., et al., Adult obesity susceptibility variants are associated with greater childhood weight gain and a faster tempo of growth: the 1946 British Birth Cohort Study. *Am J Clin Nutr*, 2012. 95(5): p. 1150–6. doi: [10.3945/ajcn.111.027870](https://doi.org/10.3945/ajcn.111.027870) PMID: [22456663](https://pubmed.ncbi.nlm.nih.gov/22456663/)
57. Sovio U., et al., Association between common variation at the FTO locus and changes in body mass index from infancy to late childhood: the complex nature of genetic association through growth and development. *PLoS Genet*, 2011. 7(2): p. e1001307. doi: [10.1371/journal.pgen.1001307](https://doi.org/10.1371/journal.pgen.1001307) PMID: [21379325](https://pubmed.ncbi.nlm.nih.gov/21379325/)
58. Graff M., et al., Genome-wide analysis of BMI in adolescents and young adults reveals additional insight into the effects of genetic loci over the life course. *Hum Mol Genet*, 2013. 22(17): p. 3597–607. doi: [10.1093/hmg/ddt205](https://doi.org/10.1093/hmg/ddt205) PMID: [23669352](https://pubmed.ncbi.nlm.nih.gov/23669352/)
59. Do R., et al., Common variants associated with plasma triglycerides and risk for coronary artery disease. *Nat Genet*, 2013. 45(11): p. 1345–52. doi: [10.1038/ng.2795](https://doi.org/10.1038/ng.2795) PMID: [24097064](https://pubmed.ncbi.nlm.nih.gov/24097064/)
60. Fromm-Dornieden C., et al., Extrinsic and intrinsic regulation of DOR/TP53NP2 expression in mice: effects of dietary fat content, tissue type and sex in adipose and muscle tissues. *Nutr Metab (Lond)*, 2012. 9(1): p. 86.
61. McCarthy M.I., Genomics, type 2 diabetes, and obesity. *N Engl J Med*, 2010. 363(24): p. 2339–50. doi: [10.1056/NEJMr0906948](https://doi.org/10.1056/NEJMr0906948) PMID: [21142536](https://pubmed.ncbi.nlm.nih.gov/21142536/)
62. Travers M.E. and McCarthy M.I., Type 2 diabetes and obesity: genomics and the clinic. *Hum Genet*, 2011. 130(1): p. 41–58. doi: [10.1007/s00439-011-1023-8](https://doi.org/10.1007/s00439-011-1023-8) PMID: [21647602](https://pubmed.ncbi.nlm.nih.gov/21647602/)
63. Li Y., et al., Genotype Imputation. *Annual Review of Genomics and Human Genetics*, 2009. 10(1): p. 387–406.
64. Marchini J., et al., A new multipoint method for genome-wide association studies by imputation of genotypes. *Nat Genet*, 2007. 39(7): p. 906–913. PMID: [17572673](https://pubmed.ncbi.nlm.nih.gov/17572673/)
65. Guan Y. and Stephens M., Practical Issues in Imputation-Based Association Mapping. *PLoS Genet*, 2008. 4(12): p. e1000279. doi: [10.1371/journal.pgen.1000279](https://doi.org/10.1371/journal.pgen.1000279) PMID: [19057666](https://pubmed.ncbi.nlm.nih.gov/19057666/)
66. Voight B.F., et al., The metabochip, a custom genotyping array for genetic studies of metabolic, cardiovascular, and anthropometric traits. *PLoS Genet*, 2012. 8(8): p. e1002793. doi: [10.1371/journal.pgen.1002793](https://doi.org/10.1371/journal.pgen.1002793) PMID: [22876189](https://pubmed.ncbi.nlm.nih.gov/22876189/)
67. Li Y., et al., MaCH: using sequence and genotype data to estimate haplotypes and unobserved genotypes. *Genetic Epidemiology*, 2010. 34(8): p. 816–34. doi: [10.1002/gepi.20533](https://doi.org/10.1002/gepi.20533) PMID: [21058334](https://pubmed.ncbi.nlm.nih.gov/21058334/)
68. Aulchenko Y.S., Struchalin M.V., and van Duijn C.M., ProbABEL package for genome-wide association analysis of imputed data. *BMC Bioinformatics*, 2010. 11: p. 134. doi: [10.1186/1471-2105-11-134](https://doi.org/10.1186/1471-2105-11-134) PMID: [20233392](https://pubmed.ncbi.nlm.nih.gov/20233392/)
69. Aulchenko Y.S., et al., GenABEL: an R library for genome-wide association analysis. *Bioinformatics*, 2007. 23(10): p. 1294–6. PMID: [17384015](https://pubmed.ncbi.nlm.nih.gov/17384015/)
70. Abecasis G.R. and Wigginton J.E., Handling marker-marker linkage disequilibrium: pedigree analysis with clustered markers. *Am J Hum Genet*, 2005. 77(5): p. 754–67. PMID: [16252236](https://pubmed.ncbi.nlm.nih.gov/16252236/)
71. Purcell S., et al., PLINK: a tool set for whole-genome association and population-based linkage analyses. *The American Journal of Human Genetics*, 2007. 81: p. 559–575. PMID: [17701901](https://pubmed.ncbi.nlm.nih.gov/17701901/)
72. Kutalik Z., et al., Methods for testing association between uncertain genotypes and quantitative traits. *Biostatistics*, 2011. 12(1): p. 1–17. doi: [10.1093/biostatistics/kxq039](https://doi.org/10.1093/biostatistics/kxq039) PMID: [20543033](https://pubmed.ncbi.nlm.nih.gov/20543033/)
73. Winkler T.W., et al., Quality control and conduct of genome-wide association meta-analyses. *Nat Protoc*, 2014. 9(5): p. 1192–212. doi: [10.1038/nprot.2014.071](https://doi.org/10.1038/nprot.2014.071) PMID: [24762786](https://pubmed.ncbi.nlm.nih.gov/24762786/)
74. Devlin B. and Roeder K., Genomic control for association studies. *Biometrics*, 1999. 55(4): p. 997–1004. PMID: [11315092](https://pubmed.ncbi.nlm.nih.gov/11315092/)
75. Willer C.J., Li Y., and Abecasis G.R., METAL: fast and efficient meta-analysis of genomewide association scans. *Bioinformatics*, 2010. 26(17): p. 2190–1. doi: [10.1093/bioinformatics/btq340](https://doi.org/10.1093/bioinformatics/btq340) PMID: [20616382](https://pubmed.ncbi.nlm.nih.gov/20616382/)
76. Benjamini Y. and Hochberg Y., Controlling the False Discovery Rate—a Practical and Powerful Approach to Multiple Testing. *Journal of the Royal Statistical Society Series B-Methodological*, 1995. 57(1): p. 289–300.



77. Rowland M.L., Self-reported weight and height. *Am J Clin Nutr*, 1990. 52(6): p. 1125–33. PMID: [2239790](#)
78. Elgar F.J., et al., Validity of self-reported height and weight and predictors of bias in adolescents. *J Adolesc Health*, 2005. 37(5): p. 371–5. PMID: [16227121](#)
79. Keith S.W., et al., Use of self-reported height and weight biases the body mass index-mortality association. *Int J Obes (Lond)*, 2011. 35(3): p. 401–8.
80. Purcell S., et al., PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet*, 2007. 81(3): p. 559–75. PMID: [17701901](#)
81. Genomes Project, C., et al., A map of human genome variation from population-scale sequencing. *Nature*, 2010. 467(7319): p. 1061–73. doi: [10.1038/nature09534](#) PMID: [20981092](#)
82. Hindorf, L.A., et al., *A Catalog of Published Genome-Wide Association Studies*. 2010.
83. Lango Allen, H., et al., Hundreds of variants clustered in genomic loci and biological pathways affect human height. *Nature*, 2010. 467(7317): p. 832–8. doi: [10.1038/nature09410](#) PMID: [20881960](#)
84. Kamatani Y., et al., Genome-wide association study of hematological and biochemical traits in a Japanese population. *Nature Genetics*, 2010. 42(3): p. 210–5. doi: [10.1038/ng.531](#) PMID: [20139978](#)
85. Franke A., et al., Genome-wide meta-analysis increases to 71 the number of confirmed Crohn's disease susceptibility loci. *Nature Genetics*, 2010. 42(12): p. 1118–25. doi: [10.1038/ng.717](#) PMID: [21102463](#)
86. Sawcer S., et al., Genetic risk and a primary role for cell-mediated immune mechanisms in multiple sclerosis. *Nature*, 2011. 476(7359): p. 214–9. doi: [10.1038/nature10251](#) PMID: [21833088](#)
87. Wang K.S., Liu X.F., and Aragam N., A genome-wide meta-analysis identifies novel loci associated with schizophrenia and bipolar disorder. *Schizophr Res*, 2010. 124(1–3): p. 192–9. doi: [10.1016/j.schres.2010.09.002](#) PMID: [20889312](#)
88. Cirulli E.T., et al., Common genetic variation and performance on standardized cognitive tests. *Eur J Hum Genet*, 2010. 18(7): p. 815–20. doi: [10.1038/ejhg.2010.2](#) PMID: [20125193](#)
89. Estrada K., et al., Genome-wide meta-analysis identifies 56 bone mineral density loci and reveals 14 loci associated with risk of fracture. *Nature Genetics*, 2012. 44(5): p. 491–501. doi: [10.1038/ng.2249](#) PMID: [22504420](#)
90. Gieger C., et al., New gene functions in megakaryopoiesis and platelet formation. *Nature*, 2011. 480(7376): p. 201–8. doi: [10.1038/nature10659](#) PMID: [22139419](#)
91. Need A.C., et al., A genome-wide study of common SNPs and CNVs in cognitive performance in the CANTAB. *Hum Mol Genet*, 2009. 18(23): p. 4650–61. doi: [10.1093/hmg/ddp413](#) PMID: [19734545](#)
92. Calvano S.E., et al., A network-based analysis of systemic inflammation in humans. *Nature*, 2005. 437(7061): p. 1032–7. PMID: [16136080](#)
93. Yang J., et al., GCTA: a tool for genome-wide complex trait analysis. *Am J Hum Genet*, 2011. 88(1): p. 76–82. doi: [10.1016/j.ajhg.2010.11.011](#) PMID: [21167468](#)
94. Kutalik Z., et al., Novel method to estimate the phenotypic variation explained by genome-wide association studies reveals large fraction of the missing heritability. *Genet Epidemiol*, 2011. 35(5): p. 341–9. doi: [10.1002/gepi.20582](#) PMID: [21465548](#)
95. Heid I.M., et al., Meta-analysis identifies 13 new loci associated with waist-hip ratio and reveals sexual dimorphism in the genetic basis of fat distribution. *Nat Genet*, 2010. 42(11): p. 949–960. doi: [10.1038/ng.685](#) PMID: [20935629](#)
96. Johnson A.D., et al., SNAP: a web-based tool for identification and annotation of proxy SNPs using HapMap. *Bioinformatics*, 2008. 24(24): p. 2938–9. doi: [10.1093/bioinformatics/btn564](#) PMID: [18974171](#)
97. Kumar P., Henikoff S., and Ng P.C., Predicting the effects of coding non-synonymous variants on protein function using the SIFT algorithm. *Nat Protoc*, 2009. 4(7): p. 1073–81. doi: [10.1038/nprot.2009.86](#) PMID: [19561590](#)
98. Wang K., Li M., and Hakonarson H., ANNOVAR: functional annotation of genetic variants from high-throughput sequencing data. *Nucleic Acids Res*, 2010. 38(16): p. e164. doi: [10.1093/nar/gkq603](#) PMID: [20601685](#)
99. Boyle A.P., et al., Annotation of functional variation in personal genomes using RegulomeDB. *Genome Res*, 2012. 22(9): p. 1790–7. doi: [10.1101/gr.137323.112](#) PMID: [22955989](#)